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# (54) Title: USE OF HEPARANASE TO IDENTIFY AND ISOLATE ANTI-HEPARANASE COMPOUND

## (57) Abstract

Purified heparanase having activity of greater than 20 units/ $\mu$ g protein, preferably greater than 50 units heparanase activity per  $\mu$ g protein, is described. The use of heparanase for screening for anti-heparanase compounds is also described. In addition, the use of the high potency heparanase to accelerate wound healing or its use as an immobilized heparanase filter connected to extracorporeal devices to degrade heparin and neutralize its anticoagulant properties during surgery is disclosed.

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7

# USE OF HEPARANASE TO IDENTIFY AND ISOLATE ANTI-HEPARANASE COMPOUND

### FIELD OF INVENTION

The present invention discloses the use of mammalian heparanase, preferably

recombinant heparanase, for screening for anti-heparanase compounds. More particularly, the present invention provides a method of selecting IHA (Inhibitors of Heparanase Activity). In addition, the present invention provides a purified heparanase, particularly suitable for use to identify and isolate anti-heparanase compounds as well as for other known uses of heparanases, such as its use to accelerate wound healing or its use as an immobilized heparanase filter

connected to extracorporeal devices to degrade heparin and neutralize its anticoagulant properties during surgery.

# **BACKGROUND OF THE INVENTION**

Elevated heparanase activity has been documented in mobile, invasive cells. Examples include; invasive melanoma, lymphoma, mastocytoma, mammary adenocarcinoma, leukemia, and rheumatoid fibroblasts. This activity has also been documented in non-pathologic situations involving the migration of lymphocytes, neutrophils, macrophages, eosinphils and platelets. An inhibitor of heparanase would therefore broadly influence the invasive potential of these diverse cells.

Inhibition of heparan sulfate degradation would also inhibit the release of bound growth factors and other biologic response modifiers that would, if released, fuel the growth of adjacent tissues and provide a supportive environment for cell growth (Rapraeger, et al., Science 252: 1705-1708, 1991). Inhibitors of heparanase activity would be of value in the treatment of arthritis, vascular restenosis, tumor growth and progression, and fibro-proliferative disorders.

Until now, the obstacles to designing a screening assay to find inhibitors of mammalian heparanase have been the unavailability of a mammalian heparanase that is purified to apparent homogeneity and the lack of information about the amino acid sequence or the 3-dimensional structure of the enzyme. Without the amino acid sequence, it has not been possible to produce recombinant mammalian heparanase to be used in large volume screening efforts. Knowledge of the tertiary and quaternary structures would facilitate rational design of IHA. This report overcomes obstacles relating to the sequence of the heparanase, and also provides a model for higher-order structure.

Heparanase refers to a mammalian enzyme which can degrade heparin proteoglycans (HPG) and/or heparan sulfate proteoglycans (HSPG).

Heparanase activity in mammalian cells is well known. It is found in various melanoma cells (Nakajima, et al., Cancer Letters 31: 277-283, 1986), mammary adenocarcinoma cells (Parish, et al., Int. J. Cancer, 40: 511-518, 1987), leukemic cells (Yahalom, et al., Leukemia

Research 12: 711-717, 1988), mast cells (Ogren and Lindahl, J. Biol. Chem. 250: 2690-2697, 1975), macrophages (Savion, et al., J. Cell. Physiol., 130: 85-92, 1987), mononuclear cells (Sewell, et al., Biochem. J. 264: 777-783, 1989), neutrophils (Matzner, et al., J. Leukocyte Biology 51: 519-524, 1992), T-cells (Vettel, et al., Eur. J. Immunol. 21: 2247-2251, 1991), platelets (Haimovitz-Friedman, et al., Blood 78: 789-796, 1991), endothelial cells (Godder, et al., J. Cell Physiol. 148: 274-280, 1991), and placenta (Klein and von Figura, BBRC 73: 569, 1976).

WO 91/02977, incorporated herein by reference, describes a substantially, but partially, purified heparanase produced by cation exchange resin chromatography and the affinity absorbent purification of heparanase-containing cell extract. WO 91/02977 also describes a method promoting wound healing utilizing compositions comprising a "purified" form of heparanase.

Others have proposed the use of purified bacterial heparanase, immobilized onto filters and connected to extracorporeal devices, to degrade heparin and neutralize its anticoagulant properties post surgery (Freed, et al., Ann. Blomed. Eng. 21: 67-76, 1993).

U.S. Patent 4,882,318 describes heparanase-inhibiting compositions for preventing tumor metastasis.

Haimovitz-Friedman et al. (Blood 78: 789-796, 1991) describe an assay for heparanase activity that involves the culturing of endothelial cells in radiolabeled <sup>35</sup>SO<sub>4</sub> to produce radiolabeled heparan sulfate proteoglycans, the removal of the cells which leaves the deposited extracellular matrix that contains the <sup>35</sup>S-HSPG, the addition of potential sources of heparanase activity, and the detection of possible activity by passing the supernatant from the radiolabeled extracellular matrix over a gel filtration column and monitoring for changes of the size of the radiolabeled material that would indicate that HSPG degradation had taken place. This assay does not have the capability for large-scale screening of inhibitors.

Nakajima et al. (Anal. Biochem. 196: 162-171, 1986) describe a solid-phase substrate for the assay of melanoma heparanase activity. Heparan sulfate from bovine lung is chemically radiolabeled by reacting it with [14C]-acetic anhydride. Free amino groups of the [14C]-heparan sulfate were acetylated and the reducing termini were aminated. The [14C]-heparan sulfate was chemically coupled to an agarose support via the introduced amine groups on the reducing termini. This substrate is limited in that it is an extensively chemically modified form of naturally occurring heparan sulfate.

Khan and Newman (Anal. Biochem. 196: 373-376, 1991) describe an indirect assay for heparanase activity. In this assay, heparin is quantitated by its ability to interfere with the color development between a protein and the dye Coomassie brilliant blue. Heparanase activity is

detected by the loss of this interference. This assay is limited in use for screening because it is so indirect that other non-heparin compounds could also interfere with the protein-dye reaction.

The CXC chemokine family (also called the intercrine α family) is one branch of the supergene "intercrine" cytokine family (Oppenheim, Ann. Rev. Bio Seam. 9: 617-648, 1991). It's members include platelet factor 4, platelet basic protein and derivatives, γIP-10, gro(α,β,γ), NAP-1/interleukin-8, mig, and ENA-78 (for review, see Miller and Krangel, Critical Reviews in Immunology 12: 17-46, 1992). The other branch, the CC chemokines or intercrine-β family, includes MIP1α, MIP1β, JE/MCP-1, RANTES, and MCAF. All members of both branches of this chemokine family characteristically are basic heparin-binding polypeptides, display molecular weights between 8 and 11 kD, share 20 - 50% homology, and function broadly in pathologic situations characterized by inflammation and tissue remodeling.

The proteolytically processed forms of platelet basic protein include CTAP-III, β-thromboglobulin, and NAP-2. β-thromboglobulin (Moore, et al., Biochim. Biophys. Acta. 379: 360-369, 1975) and CTAP-III (Castor, et al., Arthritis Rheum. 20: 859-868, 1977), were originally isolated from activated supernatants or lysates from outdated planets. Using the techniques of subcellular fractionation and radioimmunoassay, β-thromboglobulin was identified as an α-granule protein that could be released upon activation (Kaplan, et al., Blood 53: 604-618, 1979). Platelet basic protein itself was later isolated from fresh platelets, megakaryocytes, and HEL cells, an immortal human erythroleukemia cell line (Holt, et al., Biochemistry 25: 1988-1996, 1986; Holt, et al., Exp. Hematol. 16: 302-306, 1988). Walz and Baggiolini isolated the processed form of NAP-2 from platelet-containing cultures of stimulated mononuclear cells (Walz, et al., J. Exp. Med. 170: 1745-1750, 1989).

Material labeled as β-thromboglobulin is commercially available from Calbiochem, San Diego, CA (Cat. # 605165), Celsus Laboratories, Cincinnati, OH (Cat. # 41705), and Haematologic Technologies, Essex Jct., VT (Cat. # HBTG-02100. The inventors have determined, by using the "Purification Assay," that the commercial preparation have heparanase activity at a level of 0.075 units/μg. This activity is below the level of 1 unit/μg needed for the screening of anti-heparanase compounds in accordance with the assay of the subject invention.

U.S. Patent 4,897,348 describes recombinant materials and methods for producing human connective tissue-activating peptide-III (CTAP-III) and analogs thereof.

Transglutaminases catalyze the posttranslational modification of proteins by transamidation of available glutamine residues. This action results primarily in the formation of epsilon-(gamma-glutamyl)lysine cross-links (Greenberg, et al., FASEB J. 5: 3071-3076, 1991). This posttranslational modification has been reported to dramatically alter the action of some small proteins. For example, a transglutaminase produces a glutamine-lysine cross-link in the 13

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kD phospholipase A<sub>2</sub> and increases its specific enzymatic activity (Cordella-Miele, et al., J. Biol. Chem. 265: 17180-17188, 1990). A transglutaminase cross-links another small molecule, interleukin-2, and converts its activity to one that is cytotoxic to mature oligodendrocytes (Eitan and Schwartz, Science 261: 106-108,1993). The glutamine-lysine cross-link in a protein would result in the loss of overall positive charge for that protein. The transglutaminases are optimally active and generally used under reducing conditions such as dithiothreitol. The concept that glutamine-lysine cross-linking alters the activity of these small proteins may be applicable to other small molecules as well.

# SUMMARY OF THE INVENTION

The present invention discloses a method of screening for compounds having antiheparanase activity (AHA compounds), i.e. inhibitors of heparanase activity (IHA), comprising
the steps of: contacting a potential AHA compound with radiolabeled heparin/heparan sulfate
and heparanase for a time and under such conditions sufficient to allow for inhibition of
heparanase activity; detecting inhibition of heparanase activity; and selecting compounds that
inhibit heparanase activity. The present invention also discloses the amino acid sequence
identity of the heparanase that has been purified to homogeneity by chromatography under
reducing conditions. Identification of the amino acid sequence of the protein which contains
heparanase activity is crucial for the production of recombinant mammalian heparanase.

# DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a purified heparanase, and a method for producing it. The heparanase so produced has an activity of greater than 20 units/ µg protein, preferably greater than 50 units heparanase activity per µg protein (1 unit = 1% cpm < 30 K/hr using the "Purification Assay" (Example 2, Part D).

In addition, the present invention provides recombinant heparanase and a means for producing it. The term "purified heparanase" as used in the specification and claims includes the recombinant heparanase as described in the subject application. The recombinant heparanase of the subject invention can be used for the same purposes and in the same manner as the purified heparanase.

The purified heparanase of the present invention has an isoelectric point of less than 5.5 (preferably about 4.8 - 5.1) and preferably is activated by treatment with transglutaminase using reducing conditions.

The recombinant heparanase of the present invention has an isoelectric point of less than 5.5 (preferably about 4.8 - 5.1), and is isolated under reducing conditions and is activated by treatment with transglutaminase.

Suitable transglutaminases that may be used for this purpose include Activated Factor XIIIa, guinea pig liver transglutaminase, epidermal transglutaminase, keratinocyte

of:

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transglutaminase, and tissue transglutaminase.

The heparanase of the present invention has the amino acid sequence (SEQ. ID. NO: 1)

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala 5 10 15

Glu Leu Arg Cys Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys 20 25 30

10 Asn Ile Gln Ser Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln 35 40 45

Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp 50 55 60

Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gin Lys Lys Leu Ala Giy 65 70 75 80

Asp Glu Ser Ala Asp 85

encoded by the cDNA sequence (SEQ ID NO: 2) of:

- 1 AACTTGGCGA AAGGCAAAGA GGAAAGTCTA GACAGTGACT TGTATGCTGA
- 51 ACTCCGCTGC ATGTGTATAA AGACAACCTC TGGAATTCAT CCCAAAAACA
- 101 TCCAAAGTTT GGAAGTGATC GGGAAAGGAA CCCATTGCAA CCAAGTCGAA
- 5 151 GTGATAGCCA CACTGAAGGA TGGGAGGAAA ATCTGCCTGG ACCCAGATGC
  - 201 TCCCAGAATC AAGAAAATTG TACAGAAAAA ATTGGCAGGT GATGAATCTG

251 CTGAT

which corresponds to the cDNA sequence and derived amino acid sequence of CTAP-III. See Wenger et al., *Blood*, 73: 1498-1503, 1989.

30 In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 3) of:

Ser Ser Thr Lys Gly Gln Thr Lys Arg Asn Leu Ala Lys Gly Lys Glu
5 10 15

35 Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu Leu Arg Cys Met Cys lle
20 25 30

Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser Leu Glu Val 35 40 45

Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile Ala Thr Leu 50 55 60

Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys
45 65 70 75 80

Lys Ile Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala Asp
85
90

encoded by the cDNA sequence (SEQ ID NO: 4) of:

- 1 TCCTCCACCA AAGGACAAAC TAAGAGAAAC TTGGCGAAAG GCAAAGAGGA
- 5 51 AAGTCTAGAC AGTGACTTGT ATGCTGAACT CCGCTGCATG TGTATAAAGA
  - 101 CAACCTCTGG AATTCATCCC AAAAACATCC AAAGTTTGGA AGTGATCGGG
  - 151 AAAGGAACCC ATTGCAACCA AGTCGAAGTG ATAGCCACAC TGAAGGATGG
  - 201 GAGGAAAATC TGCCTGGACC CAGATGCTCC CAGAATCAAG AAAATTGTAC
  - 251 AGAAAAATT GGCAGGTGAT GAATCTGCTG AT
- which corresponds to the cDNA sequence and derived amino acid sequence of platelet basic protein. See Wenger et al., *Blood*, 73: 1498-1503, 1989 as well as Walz and Baggiolini, *BBRC* 159: 969-981, 1989; Castor, *et al.*, *BBRC* 163: 1071-1078, 1989.

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 5) of:

- 15 Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu Leu Arg Cys
  1 5 10 15
  - Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser 20 25 30

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Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile 35 40 45

Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro 25 50 55 60

Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala 65 70 75 80

30 Asp

encoded by the cDNA sequence (SEQ ID NO: 6) of:

- 1 GGCAAAGAGG AAAGTCTAGA CAGTGACTTG TATGCTGAAC TCCGCTGCAT
- 51 GTGTATAAAG ACAACCTCTG GAATTCATCC CAAAAACATC CAAAGTTTGG
- 101 AAGTGATCGG GAAAGGAACC CATTGCAACC AAGTCGAAGT GATAGCCACA
- 35 151 CTGAAGGATG GGAGGAAAAT CTGCCTGGAC CCAGATGCTC CCAGAATCAA 201 GAAAATTGTA CAGAAAAAAT TGGCAGGTGA TGAATCTGCT GAT

which corresponds to the cDNA sequence and derived amino acid sequence  $\beta$ -thromboglobulin ( $\beta$ TG). See Wenger et al., *Blood*, 73: 1498-1503, 1989.

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 7) of:

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Glu Leu Arg Cys Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys
1 5 10 15

Asn Ile Gln Ser Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln 20 25 30

Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp 35 40 45

10 Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys Leu Ala Gly
50 55 60

Asp Glu Ser Ala Asp

65 15

encoded by the cDNA sequence (SEQ ID NO: 8) of:

- 1 GAACTCCGCT GCATGTGTAT AAAGACAACC TCTGGAATTC ATCCCAAAAA
- 51 CATCCAAAGT TTGGAAGTGA TCGGGAAAGG AACCCATTGC AACCAAGTCG
- 101 AAGTGATAGC CACACTGAAG GATGGGAGGA AAATCTGCCT GGACCCAGAT
- 151 GCTCCCAGAA TCAAGAAAAT TGTACAGAAA AAATTGGCAG GTGATGAATC
  201 TGCTGAT

which corresponds to the cDNA sequence and derived amino acid sequence of neutrophil activating peptide-2.

The foregoing amino acid sequences correspond to the products of a single gene called platelet basic protein (Walz and Baggiolini, *BBRC* 159: 969-981, 1989; Castor, *et al.*, *BBRC* 163: 1071-1078, 1989). The complete gene sequence of platelet basic protein is well known. See, for example, Wenger et al., *Blood*, 73: 1498-1503, 1989 and *Proc. Natl. Acad. Sci. USA*, 90, 3660-3664, 1993.

The present invention also provides heparanase having the amino acid sequences of
other members of the CXC chemokine family [including Platelet factor 4 (SEQ. ID NO. 12),
γIP-10 (SEQ. ID NO. 14), gro/MGSA (SEQ. ID NO. 16), gro-β/MIP-2α (SEQ. ID NO. 18),
gro-γ/MIP-2β (SEQ. ID NO. 20), Interleukin-8/NAP-1 (SEQ. ID NO. 22) and ENA-78 (SEQ. ID
NO. 24)] as well as members of the CC chemokine family [including MIP-1α (SEQ. ID NO.
26), MIP-1β (SEQ. ID NO. 28), I-309 (SEQ. ID NO. 23), MCP-1 (SEQ. ID NO. 32), MCP-3
(SEQ. ID NO. 34), RANTES (SEQ. ID NO. 36), fic (SEQ. ID NO. 38) and MCP-2 (SEQ. iD
NO. 40)]; purified to apparent homogeneity, prepared in the presence of reducing conditions,
and activated by treatment with transglutaminase. Suitable transglutaminases that may be used
for this purpose include Activated Factor XIIIa, guinea pig liver transglutaminase, epidermal
transglutaminase, keratinocyte transglutaminase, and tissue transglutaminase.

In another aspect, the present invention provides a heparanase having the amino acid

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sequence (SEQ ID NO: 12) of:

Met Ser Ser Ala Ala Gly Phe Cys Ala Ser Arg Pro Gly Leu Leu Phe Leu Gly Leu Leu Leu Leu Pro Leu Val Val Ala Phe Ala Ser Ala Glu Ala Glu Glu Asp Gly Asp Leu Gln Cys Leu Cys Val Lys Thr Thr Ser Gln Val Arg Pro Arg His Ile Thr Ser Leu Glu Val Ile Lys Ala Gly Pro His Cys Pro Thr Ala Gln Leu Ile Ala Thr Leu Lys Asn Gly Arg Lys Ile Cys Leu Asp Leu Gln Ala Pro Leu Tyr Lys Lys Ile Ile Lys Lys Leu Glu Ser

encoded by the cDNA sequence (SEQ ID NO: 13) of:

- 1 CCGCAGCATG AGCTCCGCAG CCGGGTTCTG CGCCTCACGC CCCGGGCTGC
- 10 51 TGTTCCTGGG GTTGCTGCTC CTGCCACTTG TGGTCGCCTT CGCCAGCGCT
  - 101 GAAGCTGAAG AAGATGGGGA CCTGCAGTGC CTGTGTGTA AGACCACCTC
  - 151 CCAGGTCCGT CCCAGGCACA TCACCAGCCT GGAGGTGATC AAGGCCGGAC
  - 201 CCCACTGCCC CACTGCCCAA CTGATAGCCA CGCTGAAGAA TGGAAGGAAA
  - 251 ATTTGCTTGG ACCTGCAAGC CCCGCTGTAC AAGAAAATAA TTAAGAAACT
  - 301 TTTGGAGAGT TAGCTACTAG CTGCCTACGT GTGTGCATTT GCTATATAGC
  - 351 ATACTTCTTT TITCCAGTTT CAATCTAACT GTGAAAGAAA CTTCTGATAT
  - 401 TTGTGTTATC CTTATGATTT TAAATAAACA AAATAAATC

which corresponds to the cDNA sequence and derived amino acid sequence of platelet factor 4. See Poncz et al., *Blood* 69, 219-223 (1987).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 14) of:

Met Asn Gin Thr Ala Ile Leu Ile Cys Cys Leu Ile Phe Leu Thr Leu Ser Gly Ile Gin Gly Val Pro Leu Ser Arg Thr Val Arg Cys Thr Cys Ile Ser Ile Ser Asn Gin Pro Val Asn Pro Val Asn Pro Arg Ser Leu Glu Lys Leu Glu Ile Ile Pro Ala Ser Gin Phe Cys Pro Arg Val Giu Ile Ile Ala Thr Met Lys Lys Gly Glu Lys Arg Cys Leu Asn Pro Glu Ser

Lys Ala Ile Lys Asn Leu Leu Lys Ala Val Ser Lys Glu Met Ser Lys Arg Ser Pro encoded by the cDNA sequence (SEQ ID NO: 15) of:

- 1 GAGACATTCC TCAATTGCTT AGACATATTC TGAGCCTACA GCAGAGGAAC
- 51 CTCCAGTCTC AGCACCATGA ATCAAACTGC GATTCTGATT TGCTGCCTTA
- 101 TCTTTCTGAC TCTAAGTGGC ATTCAAGGAG TACCTCTCTC TAGAACCGTA
- 151 CGCTGTACCT GCATCAGCAT TAGTAATCAA CCTGTTAATC CAAGGTCTTT
- 201 AGAAAAACTT GAAATTATTC CTGCAAGCCA ATTTTGTCCA CGTGTTGAGA
- 251 TCATTGCTAC AATGAAAAAG AAGGGTGAGA AGAGATGTCT GAATCCAGAA
- 301 TCGAAGCCA TCAAGAATTT ACTGAAAGCA GTTAGCAAGG AAATGTCTAA
- 35 351 AAGATCTCCT TAAAACCAGA GGGGAGCAAA ATCGATGCAG TGCTTCCAAG
  - 401 GATGGACCAC ACAGAGGCTG CCTCTCCCAT CACTTCCCTA CATGGAGTAT

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- 451 ATGTCAAGCC ATAATTGTTC TTAGTTTGCA GTTACACTAA AAGGTGACCA
- 501 ATGATGGTCA CCAAATCAGC TGCTACTACT CCTGTAGGAA GGTTAATGTT
- 551 CATCATCCTA AGCTATTCAG TAATAACTCT ACCCTGGCAC TATAATGTAA
- 601 GCTCTACTGA GGTGCTATGT TCTTAGTGGA TGTTCTGACC CTGCTTCAAA
- 5 which corresponds to the cDNA sequence and derived amino acid sequence γIP-10. See Luster et al., Nature 315, 672-676 (1985).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 16) of:

Met Ala Arg Ala Ala Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu Arg Val Ala Leu Leu Leu Leu Leu Leu Leu Val Ala Ala Gly Arg Arg Ala Ala Gly Ala Ser Val Ala Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr Leu Gln Gly Ile His Pro Lys Asn Ile Gln Ser Val Asn Val Lys Ser Pro Gly Pro His Cys Ala Gln Thr Glu Valle Ala Thr Leu Lys Asn Gly Arg Lys Ala Cys Leu Asn Pro Ala Ser Pro Ile Val Lys Lys Ile Ile Glu Lys Met Leu Asn Ser Asp Lys Ser Asn

- 15 encoded by the cDNA sequence (SEQ ID NO: 17) of:
  - 1 CTCGCCAGCT CTTCCGCTCC TCTCACAGCC GCCAGACCCG CCTGCTGAGC
  - 51 CCCATGGCC GCGCTGCTCT CTCCGCCGC CCCAGCAATC CCCGGCTCCT
  - 101 GCGAGTGGCA CTGCTGCTCC TGCTCCTGGT AGCCGCTGGC CGGCGCGCAG
  - 151 CAGGAGCGTC CGTGGCCACT GAACTGCGCT GCCAGTGCTT GCAGACCCTG
  - 201 CAGGGAATTC ACCCCAAGAA CATCCAAAGT GTGAACGTGA AGTCCCCCGG
  - 251 ACCCCACTGC GCCCAAACCG AAGTCATAGC CACACTCAAG AATGGGCGGA
  - 301 AAGCTTGCCT CAATCCTGCA TCCCCCATAG TTAAGAAAAT CATCGAAAAG
  - 351 ATGCTGAACA GTGACAAATC CAACTGACCA GAAGGGAGGA GGAAGCTCAC
  - 401 TGGTGGCTGT TCCTGAAGGA GGCCCTGCCC TTATAGGAAC AGAAGAGGAA
- 25 451 AGAGAGACAC AGCTGCAGAG GCCACCTGGA TTGTGCCTAA TGTGTTTGAG
  - 501 CATCGCTTAG GAGAAGTCTT CTATTTATTT ATTTATTCAT TAGTTTTGAA
  - 551 GATTCTATGT TAATATTTTA GGTGTAAAAT AATTAAGGGT ATGATTAACT
  - 601 CTACCTGCAC ACTGTCCTAT TATATTCATT CTTTTTGAAA TGTCAACCCC
  - 651 AAGTTAGTTC AATCTGGATT CATATTTAAT TTGAAGGTAG AATGTTTTTA
- 30 701 AATGITCTCC AGTCATTATG TTAATATTTC TGAGGAGCCT GCAACATGCC
  - 751 AGCCACTGTG ATAGAGGCTG GCGGATCCAA GCAAATGGCC AATGAGATCA
  - 801 TTGTGAAGGC AGGGGAATGT ATGTGCACAT CTGTTTTGTA ACTGTTTAGA
  - 851 TGAATGTCAG TTGTTATTTA TTGAAATGAT TTCACAGTGT GTGGTCAACA
  - 901 TTTCTCATGT TGAAACTTTA AGAACTAAAA TGTTCTAAAT ATCCCTTGGA
- 35 951 CATTITATGT CITTCTTGTA AGGCATACTG CCTTGTTTAA TGGTAGTTTT
  - 1001 ACAGTGTTTC TGGCTTAGAA CAAAGGGGCT TAATTATTGA TGTTTTCGGA

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which corresponds to the cDNA sequence and derived amino acid sequence of gro/MGSA (melanoma growth stimulatory activity). See Anisowicz et al., Proc. Natl. Acad. Sci. U.S.A. 84, 7188-7192 (1987).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 18) of:

Met Ala Arg Ala Thr Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu Arg Val Ala Leu Leu Leu Leu Leu Leu Und Ala Ala Ser Arg Arg Ala Ala Gly Ala Pro Lys Ala Thr Glu Lys Arg Cys Gln Cys Lys Gln Thr Leu Gln Gly Ile His Leu Lys Asn Ile Gln Ser Val Lys Val Lys Ser Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn Gly Gln Lys Ala Cys Leu Asn Pro Ala Ser Pro Met Val Lys Lys Ile Ile Glu Lys Met Leu Lys

encoded by the cDNA sequence (SEQ ID NO: 19) of:

- 1 CTCTCCTCCT CGCACAGCCG CTCGAACCGC CTGCTGAGCC CCATGGCCCG
- 51 CGCCACGCTC TCCGCCGCCC CCAGCAATCC CCGGCTCCTG CGGGTGGCGC
- 15 101 TGCTGCTCCT GCTCCTGGTG GCCGCCAGCC GGCGCGCAGC AGGAGCGCCC
  - 151 CTGGCCACTG AACTGCGCTG CCAGTGCTTG CAGACCCTGC AGGGAATTCA
  - 201 CCTCAAGAAC ATCCAAAGTG TGAAGGTGAA GTCCCCCGGA CCCCACTGCG
  - 251 CCCAAACCGA AGTCATAGCC ACACTCAAGA ATGGGCAGAA AGCTTGTCTC
  - 301 AACCCCGCAT CGCCCATGGT TAAGAAAATC ATCGAAAAGA TGCTGAAAAA
  - 351 TGGCAAATCC AACTGACCAG AAGGAAGGAG GAAGCTTATT GGTGGCTGTT
  - 401 CCTGAAGGAG GCCCTGCCCT TACAGGAACA GAAGAGGAAA GAGAGACACA
  - 451 GCTGCAGAGG CCACCTGGAT TGCGCCTAAT GTGTTTGAGC ATCACTTAGG
  - 501 AGAAGTCTTC TATTTATTTA TITATTTATT TATTTGTTTG TITTAGAAGA
  - 551 TTCTATGTTA ATATTTTATG TGTAAAATAA GGTTATGATT GAATCTACTT
  - 601 GCACACTCTC CCATTATATT TATTGTTTAT TTTAGGTCAA ACCCAAGTTA
    - 651 GTTCAATCCT GATTCATATT TAATTTGAAG ATAGAAGGTT TGCAGATATT
    - 701 CTCTAGTCAT TTGTTAATAT TTCTTCGTGA TGACATATCA CATGTCAGCC
    - 751 ACTGTGATAG AGGCTGAGGA ATCCAAGAAA ATGGCCAGTG AGATCAATGT
    - 801 GACGGCAGGG AAATGTATGT GTGTCTATTT TGTAACTGTA AAGATGAATG
- 30 851 TCAGTTGTTA TTTATTGAAA TGATTTCACA GTGTGTGGTC AACATTTCTC
  - 901 ATGTTGAAGC TTTAAGAACT AAAATGTTCT AAATATCCCT TGGACATTTT
  - 951 ATGTCTTTCT TGTAAGGCAT ACTGCCTTGT TTAATGTTAA TTATGCAGTG
  - 1001 TITCCCTCTG TGTTAGAGCA GAGAGGTTTC GATATTTATT GATGTTTTCA
  - 1051 CAAAGAACAG GAAAATAAAA TATTTAAAAA T
- which corresponds to the cDNA sequence and derived amino acid sequence gro-β/MIP-2α (macrophage inflammatory protein 2-α). See Tekamp-Olson et al., J. Exp. Med. 172, 911-919

(1990).

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In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 20) of:

Met Ala His Ala Thr Leu Ser Ala Ala Pro Ser Asn Pro Arg Leu Leu Arg Val Ala Leu Leu Leu Leu Leu Val Ala Ala Ser Arg Arg Ala Ala Gly Ala Ser Val Val Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr Leu Gln Gly Ile His Leu Lys Asn Ile Gln Ser Val Asn Val Arg Ser Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn Gly Lys Lys Ala Cys Leu Asn Pro Ala Ser Pro Met Val Gln Lys Ile Ile Glu Lys Ile Leu Asn Lys Gly Ser Thr Asn

- 10 encoded by the cDNA sequence (SEQ ID NO: 21) of:

  - 51 TCTCCGCCGC CCCCAGCAAT CCCCGGCTCC TGCGGGTGGC GCTGCTGCTC
  - 101 CTGCTCCTGG TGGCCGCCAG CCGGCGCGCA GCAGGAGCGT CCGTGGTCAC
  - 151 TGAACTGCGC TGCCAGTGCT TGCAGACACT GCAGGGAATT CACCTCAAGA
- 15 201 ACATCCAAAG TGTGAATGTA AGGTCCCCCG GACCCCACTG CGCCCAAACC
  - 251 GAAGTCATAG CCACACTCAA GAATGGGAAG AAAGCTTGTC TCAACCCCGC
  - 301 ATCCCCCATG GTTCAGAAAA TCATCGAAAA GATACTGAAC AAGGGGAGCA
  - 351 CCAACTGACA GGAGAGAAGT AAGAAGCTTA TCAGCGTATC ATTGACACTT
  - 401 CCTGCAGGGT GGTCCCTGCC CTTACCAGAG CTGAAAATGA AAAAGAGAAC
- 20 451 AGCAGCTTTC TAGGGACAGC TGGAAAGGAC TTAATGTGTT TGACTATTTC
  - 501 TTACGAGGGT TCTACTTATT TATGTATTTA TTTTTGAAAG CTTGTATTTT
  - 551 AATATITTAC ATGCTGTTAT TTAAAGATGT GAGTGTGTTT CATCAAACAT
  - 601 AGCTCAGTCC TGATTATTTA ATTGGAATAT GATGGGTTTT AAATGTGTCA
  - 651 TTAAACTAAT ATITAGTGGG AGACCATAAT GTGTCAGCCA CCTTGATAAA
- 25 701 TGACAGGGTG GGGAACTGGA GGGTGGGGGG ATTGAAATGC AAGCAATTAG
  - 751 TGGATCACTG TTAGGGTAAG GGAATGTATG TACACATCTA TITTTTATAC
  - 801 TTTTTTTTA AAAAAAGAAT GTCAGTTGTT ATTTATTCAA ATTATCTCAC
  - 851 ATTATGTGTT CAACATTTTT ATGCTGAAGT TTCCCTTAGA CATTTTATGT
  - 901 CTTGCTTGTA GGGCATAATG CCTTGTTTAA TGTCCATTCT GCAGCGTTTC
- 30 951 TCTTTCCCTT GGAAAAGAGA ATTTATCATT ACTGTTAC

which corresponds to the cDNA sequence and derived amino acid sequence *gro-γ*/MIP-2β (macrophage inflammatory protein 2-β). See Tekamp-Olson et al., J. Exp. Med. 172, 911-919 (1990).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 22) of:

Met Thr Ser Lys Leu Ala Val Ala Leu Leu Ala Ala Phe Leu Ile Ser Ala Ala Leu Cys

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Glu Gly Ala Val Leu Pro Arg Ser Ala Lys Glu Leu Arg Cys Gln Cys Ile Lys Thr Tyr Ser Lys Pro Phe His Pro Lys Phe Ile Lys Glu Leu Arg Val Ile Glu Ser Gly Pro His Cys Ala Asn Thr Glu Ile Ile Val Lys Leu Ser Asp Gly Arg Glu Leu Cys Leu Asp Pro Lys Glu Asn Trp Val Gln Arg Val Val Glu Lys Phe Leu Lys Arg Ala Glu

- 5 encoded by the cDNA sequence (SEQ ID NO: 23) of:
  - 1 ATGACTTCCA AGCTGGCCGT GGCTCTCTTG GCAGCCTTCC TGATTTCTGC
  - 51 AGCTCTGTGT GAAGGTGCAG TTTTGCCAAG GAGTGCTAAA GAACTTAGAT
  - 101 GTCAGTGCAT AAAGACATAC TCCAAACCTT TCCACCCCAA ATTTATCAAA
  - 151 GAACTGAGAG TGATTGAGAG TGGACCACAC TGCGCCAACA CAGAAATTAT
  - 201 TGTAAAGCTT TCTGATGGAA GAGAGCTCTG TCTGGACCCC AAGGAAAACT
  - 251 GGGTGCAGAG GGTTGTGGAG AAGTTTTTGA AGAGGGCTGA G which corresponds to the cDNA sequence and derived amino acid sequence Interleukin-8/NAP-1 (neutrophil activating protein-1). See Kunser et al., Kidney Int. 39, 1240-1248 (1991).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 24) of:

Ala Gly Pro Ala Ala Ala Val Leu Arg Glu Lys Arg Cys Val Cys Leu Gln Thr Thr Gln Gly Val His Pro Lys Met Ile Ser Asn Leu Gln Val Phe Ala Ile Gly Pro Gln Cys Ser Lys Val Glu Val Val Ala Ser Leu Lys Asn Gly Lys Glu Ile Cys Leu Asp Pro Glu Ala Pro Phe Leu Lys Lys Val Ile Gln Lys Ile Leu Asp Gly Gly Asn Lys Glu Asn encoded by the cDNA sequence (SEQ ID NO: 25) of:

- 1 GTGTTGCGGG AACTGCGGTG CGTGTTTTA CAGACCACGC AGGGAGTTCA
- 51 TCCCAAAATG ATCAGTAATC TGCAAGTGTT CGCCATAGGC CCACAGTGCT
- 101 CCAAGGTGGA AGTGGTAGCC TCCCTGAAGA ACGGGAAGGA AATTTGTCTT
- 151 GATCCAGAAG CCCCTTTTCT AAAGAAAGTC ATCCAGAAAA TCCTCGACGG
- 25 201 CGGCAACAAA GAAAAC

which corresponds to the cDNA sequence and derived amino acid sequence of a novel inflammatory peptide (ENA-78) with homology to interleukin 8. See Walz et al., J. Exp. Med. 174, 1355-1362 (1991).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 26) of:

Met Gln Val Ser Thr Ala Ala Leu Ala Val Leu Cys Thr Met Ala Leu Cys Asn Gln Val Leu Ser Ala Pro Leu Ala Ala Asp Thr Pro Thr Ala Cys Cys Phe Ser Tyr Thr Ser Arg Gln Ile Pro Gln Asn Phe Ile Ala Asp Tyr Phe Glu Thr Ser Ser Gln Cys Ser Lys Pro Ser Val Ile Phe Leu Thr Lys Arg Gly Arg Gln Val Cys Ala Asp Pro Ser Glu Glu Trp Val Gln Lys Tyr Val Ser Asp Leu Glu Leu Ser Ala

encoded by the cDNA sequence (SEQ ID NO: 27) of:

1 GAATTCAAGG CCTGTCCTGG TTTGGTCCCA ATTTACCTTT ATCATCCATA 51 TTCACCCCCA CTGCTCTGCA GCTCCACTGA AGCACCCCCT CTTTCCTCTG 101 AGCCACAATG TCACACCCAG GACTCTGCCT CAGCTGGGCC TCCACTGCCC 151 ACCCATCTAT AGATGCCTAA ATCCCGGGCA GTTATCCAGA CACAACTAAA 201 GTTCCATCCC TTCCATGAAG CCTTCCCCAA CCCTCTGGTG GAAGGTCACT 251 TCTTCCTCAT GGGGTTCTGA GCTTTCATTT CTTTTTCTAO TAAGAGTTTT 301 ACAATTACCT GTTCATACAC TCTACCTGCC CCCATGAGAC CAGGGGCATC 351 TCAGAAACAA AGATCATTAA AACCAACTAA ATCTATTTCT CATTATAAAA 401 TGAGATATGC TGATTGATTG CAAAATAATA AAATAACAAA GTATGGAAAA 451 GAAAAAAAA AGCATATAAT CTGGCTGAGA AGGTAGAGAC CCTTCCACAC 10 501 CACTGAAATT ATGTGTTGAA AAGAATAAGG AAAAAACTGC TTCAGTTTGG 551 CATTATITAT GTAAGTATAG TATAGGATCC TTAAAATGGT TCAAAGAAAT 601 GGGAAATCAA GACTTCATTT TGGCAAAGCC ATTGAACAGA AACTGTAGCA 651 TATITATCAG TAATITCTTT CAGATTAAAC AACTGACAAC AACCCACTTT 701 TCAACCAGTG ATGTTGGAAA TGTTTTAAAA CAAAATTAGT TCATAAATTT 15 751 GTGGGTTGAC CAAGAAGGTA ATAAAGTCTC ACTAAATAAA ATGAGGAAAA 801 TTCAGAAAAA GAAAAAATA AGAAAATAAA TCACCCATGG ATCTAAGCAC 851 TATTCATTCT TTAAGGCATG TATTTCCAAG CCTTTTAATT TTTTCATGCC 901 TAGAGTTGGC ATGGCATATA TATATCTTTA TACAATTCTT CAAATTTTAT 951 AGAATITGTA TAATGTTTTA TCTTGCTTTT TTTTTAACCA CTGATGTTAT 20 1001 AAGCATATTT ATGCCACTTC ATTCACGTTA GAGACTTAAT AATAAAGGAT 1051 CTTGTGGATA ATTTATCATT CCCTGATAGA GAAAAATTTA GCTTTGCTTA 1101 TTTTAGAGTT ATAAATGATG CTGGGTCAGG TATCTTTATG TTTGAAGATG 1151 GCTCCATATT TGGGTTGTTT CCACAGAACT CTTTCCAGAA ATGCTTTTTC 1201 TAGGTTAATG GCTACACATA TTTCTAGGCA CCTGACATAC TGACACCCAC 25 1251 CTCTAAAGTA TTTTTATGAT CCACAACTAG CGTTTAACAC AGCGCCCCAG 1301 TCACTCCGAG ACTAATAAAT AGACAAATGA CTGAAACGTG ACCTCATGCT 1351 TTCTATTCCT CCAGCTTTCA TTGAGTTCCT TTCCTCTGGG AGGACTGGGG 1401 GTTGTCTAGC CCTCCACAGC ATCAGCCCAT TGACCCTATC CTTGTGGTTA 1451 TAGCAGCTGA GGAAGCAGAA TT: AGCTCT GTGGGAAGGA ATGGGGCTGG 30 1501 AGAGTTCATG CATAGACCAA TTCTTTTTT TTTTTTTTT TGAGATGGAG 1551 TTTCACTTTT GTTGCCCAGG CTGGAGTGCA ATGGCATGAT CTCAGCTCAC 1601 CACAGCCCC ACCTCCTGGG TTCAAGCGAT TCTCCTGCCC TCAGCCTCCC 1651 GAGTAGCTGG GATTACAGGC ATGTGCCACC ACGCCTGACT ACTTTTGTAT 1701 TTTTAGTAGA GATGGAGTTT CTCTTTCTTG GTCAGGTTGG TCTCAAACTC 35 1751 CTGACCTCAG GTGATCTGCA GCCTCGGCCT CCAAAGTGTT GGGATTACAG

WO 95/04158 PCT/US94/08207

	1801	GTGTGAGCGA CCATGCCTGG CTGCATAGAC CAGTTCTTAT GAGAAGGGAT
	1851	CAACTAAGAA TAGCCTTGGG TTGACACACA CCCCTCTTCA CACTCACAGG
	1901	AGAAACCCCA TGAAGCTAGA ACCAGTCATG AGTTGAGAGC TGAGAGTTAG
	1951	AGAGTAGCTC AGAGATGCTA TTCTTGGATA TCCTGAGCCC CTGTGGTCAC
5	2001	CAGGGACCCT GAGTTGTGCA ACACTCAGCA TGACAGCATC ACTACACTTA
	2051	AAAATTTCCC TCCTCACCCC CAGATTCCAT TTCCCCATCC GCCAGGGCTG
	2101	CCTATAAAGA GGAGAGATGG CTTCAGACAT CAGAAGGACG CAGGCAGCAA
	2151	AGAGTAGTCA GTCCCTTCTT GGCTCTGCTG ACACTCGAGC CCACATTCCA
	2201	TCACCTGCTC CCAATCATGC AGGTCTCCAC TGCTGCCCTT GCCGTCCTCC
10	2251	TCTGCACCAT GGCTCTCTGC AACCAGGTCC TCTCTGCACC ACGTGAGTCC
	2301	ATGTTGTTGT TGTGGGTATC ACCACTCTCT GGCCATGGTT AGACCACATC
	2351	AGTCTTTTT TGTGGCGTGA GAGGCCCCGA AGAGAAAGA AGGAAGTTCT
	2401	TAAAGCGCTG CCAAACACCT TGGTCTTTTT CTTCACAACT TTTATTTTTA
	2451	TCTCTAGAAG GGGTCTTAGC CCTCCTAGTC TCCAGGTATG AGAATCTAGG
15	2501	CAGGGGCAGG GGAGTTACAG TCCCTTGTAC AGATAGAAAA ACAGGGTTCA
	2551	AAACGAATCA GTTTGCAAGA GGCAGAATCC AGGGCTGCTT ACTTCCCAGT
	2601	GGGGTCTGTT CTTCACTCTC CAGCTCACCC TAGTCTCCCA GGAGCCCTGT
	2651	CCCTTGGATG TCTTATGAGA GATGTCCAGG GCTTCTCTTG GGCTGGGGTA
	2701	TGACTTCTTG AACCGACAAA ATTCCATGAA GAGAGCTAAG AGAACAGTCC
20	2751	ATTCAGGTAT CTGGATCACA TAGAGAAACA GAGAACCCAC TATGAAGAGT
	2801	CAAGGGAAA GAGGAATATA GACAGAAACA AAGAGACATT TCTCTGCAA
	2851	ACCCCCAAA TGCCTTGCAG TCACTTGGTC TGAGCAAGCC TGCCCTCCTC
	2901	AACCACTCAG GGATCAGAAG CTGCCTGGCC TTTTCTTCTG AGCTGTGACT
	2951	TGGGCTTATT CTCTCCTTTC TCCGCAGTTG CTGCTGACAC GCCGACCGCC
25	3001	TGCTGCTTCA GCTACACCTC CCGACAGATT CCACAGAATT TCATAGCTGA
	3051	CTACTTTGAG ACGAGCAGCC AGTGCTCCAA GCCCAGTGTC ATGTAAGTGC
	3101	CAGTCTTCCT GCTCACCTCT AGGGAGGTAG GGAGTGTCAG GGTGGGGGCA
		GAAACAGGCC AGAAGGCCAT CCTGGAAAGG CCCAGCCTTC AGGAGCCTAT
	3201	CGGGGATACA GGACGCAGGG CACTGAGGTG TGACCTGACT TGGGGCTGGA
30	3251	GTGAGGTGGG TGTTACAGAG TCAGGAAGGG CTGCCCCAGG CCAGAGGAAA
	3301	GGGACAGGAA GAAGGAGGCA GCAGGACACT CTGAGGGCCC CCTTGCCTGC
	3351	AGTCACTGAG AGAAGCTCTC TAGACGGAGA TAGGCAGGGG GCCCCTGAGA
	3401	GAGGAGCAGG CCTTGAGCTG CCCAGGACAG AGAGCAGGAT GTCAGGGCCA
	3451	TGGTGGGCCC AGGATTCCCC GGCTGGATTC CCCAGTGCTT AACTCTTCCT
35	3501	CCCTTCTCCA CAGCTTCCTA ACCAAGAGAG GCCGGCAGGT CTGTGCTGAC
	3551	CCCAGTGAGG AGTGGGTCCA GAAATACGTC AGTGACCTGG AGCTGAGTGC

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	3601	CTGAGGGGTC CAGAAGCTTC GAGGCCCAGC GACCTCAGTG GGCCCAGTGC
	3651	GGAGGAGCAG GAGCCTGAGC CTTGGGAACA TGCGTGTGAC CTCCACAGCT
	3701	ACCTCTTCTA TGGACTGGTT ATTGCCAAAC AGCCACACTG TGGGACTCTT
	3751	CTTAACITAA ATTTTAATTT ATTTATACTA TITAGTTTTT ATAATTTATT
5	3801	TTTGATTTCA CAGTGTGTTT GTGATTGTTT GCTCTGAGAG TTCCCCCTGT
	3851	CCCTCCACC TTCCCTCACA GTGTGTCTGG TGACAACCGA GTGGCTGTCA
	3901	TCGGCCTGTG TAGGCAGTCA TGGCACCAAA GCCACCAGAC TGACAAATGT
	3951	GTATCAGATG CTTTTGTTCA GGGCTGTGAT CGGCCTGGGC AAATAATAAA
	4001	GATGTTCTTT TAAACGGTAA ACCAGTATTG AGTTTGGTTT TGTTTTTCTG
10	4051	GCAAATCAAA ATCACTGGTT AAGAGGAATC ATAGGCAAAG ATTAGGAAGA
	4101	GGTGAAATGG AGGGAAATTG GGAGAGATGG GGAGCGCTGC GACAGAGTT
	4151	TOCACTTCAC AAAATIICTGG AACATITGAAA CTACGAATAT GITTATAACTG
	4201	AAATCGTAAT ATGCACGCTC TAGGAGAATT AACTACTTGA ATGGCCAC
	4251	TTAAGCAGAG TATTCTGTAG GGCATATTCA TGATGAATCA AGCTCTTAAT
15	4301	AGCAATTATT TACATTGTTG AGGCTTACTC CTCCTACTGA GTGCTTTTTA
	4351	TACATTGTTC ATTTAATCTT ACCAATGCAA TAGTACAGCT TAGGTACTAT
	4401	TAATACCTCC ACTTGACAGA AAAGTAACCC AGGGCTCAGA AAGGTTAGAC
	4451	AACTTGGCTG AGGTTACACA GCACGTAAAC GGTCAATTGT GTTCCAAAAC
	4501	TGGACTTTTA TTGAACTACA GACTATGCTG TTAACCATTG ACCAAGTTAT
20	4551	TTCCCAAAGT ATGACCCGCC TATACTCAAA TCTTACCCCA TTCTTTAACA
	4601	GATGATACTT TATCCATTGC AACCACTTCC TGTCAGGATT CTGAGTTGAC
	4651	ATAGAGTGTT TCAGCAGTGA TTATTTAAGC CAATTACATC AGGATCTTTA
	4701	GGTGTAGACC TGGGAACTGA TATTTTTATC AAGCTCATGA GGTGTTCCAT
	4751	AGCATGTTAA TGACTGAGAG CCACTGTCAA TAGAATTC

which corresponds to the cDNA sequence and derived amino acid sequence MIP-1α (macrophage inflammatory protein 1-α). See Blum et al., DNA Cell Biol. 9, 589-602 (1990).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 28) of:

Met Lys Leu Cys Val Thr Val Leu Ser Leu Leu Met Leu Val Ala Ala Phe Cys Ser Pro Ala Leu Ser Ala Pro Met Gly Ser Asp Pro Pro Thr Ala Cys Cys Phe Ser Tyr Thr Ala Arg Lys Leu Pro Arg Asn Phe Val Val Asp Tyr Tyr Glu Thr Ser Ser Leu Cys Ser Gln Pro Ala Val Val Phe Gln Thr Lys Arg Ser Lys Gln Val Cys Ala Asp Pro Ser Glu Ser Trp Val Gln Glu Tyr Val Tyr Asp Leu Glu Leu Asn

encoded by the cDNA sequence (SEQ ID NO: 29) of:

- 1 TTCCCCCCC CCCCCCCC CCCGCCGA GCACAGGACA CAGCTGGGTT
- 51 CTGAAGCTTC TGAGTTCTGC AGCCTCACCT CTGAGAAAAC CTCTTTTCCA

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- 101 CCAATACCAT GAAGCTCTGC GTGACTGTCC TGTCTCTCCT CATGCTAGTA
- 151 GCTGCCTTCT GCTCTCCAGC GCTCTCAGCA CCAATGGGCT CAGACCCTCC
- 201 CACCGCCTGC TGCTTTTCTT ACACCGCGAG GAAGCTTCCT CGCAACTTTG
- 251 TGGTAGATTA CTATGAGACC AGCAGCCTCT GCTCCCAGCC AGCTGTGGTA
- 301 TTCCAAACCA AAAGAAGCAA GCAAGTCTGT GCTGATCCCA GTGAATCCTG
  - 351 GGTCCAGGAG TACGTGTATG ACCTGGAACT GAACTGAGCT GCTCAGAGAC
  - 401 AGGAAGTCTT CAGGGAAGGT CACCTGAGCC CGGATGCTTC TCCATGAGAC
  - 451 ACATCTCCTC CATACTCAGG ACTCCTCTCC GCAGTTCCTG TCCCTTCTCT
  - 501 TAATTTAATC TTTTTTATGT GCCGTGTTAT TGTATTAGGT GTCATTTCCA
- 551 TTATTTATAT TAGTTTAGCC AAAGGATAAG TGTCCTATGG GGATGGTCCA
  - 601 CTGTCACTGT TTCTCTGCTG TTGCAAATAC ATGGATAACA CATTTGATTC
- 651 TGTGTGTTTT CCATAATAAA ACTTTAAAAT AAAATGCAGA CAGTTA which corresponds to the cDNA sequence and derived amino acid sequence MIP-1β (macrophage inflammatory protein 1-β). See Lipes et al., Proc. Natl. Acad. Sci. U.S.A. 85, 9704-9708 (1988).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 30) of:

Met Gln Ile Ile Thr Thr Ala Leu Val Cys Leu Leu Leu Ala Gly Met Trp Pro Glu Asp Val Asp Ser Lys Ser Met Gln Val Pro Phe Ser Arg Cys Cys Phe Ser Phe Ala Glu Gln Glu Ile Pro Leu Arg Ala Ile Leu Cys Tyr Arg Asn Thr Ser Ser Ile Cys Ser Asn Glu Gly Leu Ile Phe Lys Leu Lys Arg Gly Lys Glu Ala Cys Ala Leu Asp Thr Val Gly Trp Val Gln Arg His Arg Lys Met Leu Arg His Cys Pro Ser Lys Arg Lys encoded by the cDNA sequence (SEQ ID NO: 31) of:

- 1 ACCAGGCTCA TCAAAGCTGC TCCAGGAAGG CCCAAGCCAG ACCAGAAGAC
- 51 ATGCAGATCA TCACCACAGC CCTGGTGTGC TTGCTGCTAG CTGGGATGTG
  - 101 GCCGGAAGAT GTGGACAGCA AGAGCATGCA GGTACCCTTC TCCAGATGTT
  - 151 GCTTCTCATT TGCGGAGCAA GAGATTCCCC TGAGGGCAAT CCTGTGTTAC
  - 201 AGAAATACCA GCTCCATCTG CTCCAATGAG GGCTTAATAT TCAAGCTGAA
  - 251 GAGAGGCAAA GAGGCCTGCG CCTTGGACAC AGTTGGATGG GTTCAGAGGC
- 30 301 ACAGAAAAT GCTGAGGCAC TGCCCGTCAA AAAGAAAATG AGCAGATTTC
  - 351 TTTCCATTGT GGGCTCTGGA AACCACATGG CTTCACCTGT CCCCGAAACT
  - 401 ACCAGCCCTA CACCATTCCT TCTGCCCTGC TTTTGCTAGG TCACAGAGGA
  - 451 TCTGCTTGGT CTTGATAAGC TATGTTGTTG CACTTTAAAC ATTTAAATTA
  - 501 TACAATCATC AACCCCCAAC
- which corresponds to the cDNA sequence and derived amino acid sequence human secreted protein (I-309). See Miller et al., J. Immunol. 143, 2907-2916 (1989).

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In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 32) of:

Met Lys Val Ser Ala Ala Leu Leu Cys Leu Leu Leu Ile Ala Ala Thr Phe Ile Pro Gln Gly Lys Ala Gln Pro Asp Ala Ile Asn Ala Pro Val Thr Cys Cys Tyr Asn Phe Thr Asn Arg Lys Ile Ser Val Gln Arg Leu Ala Ser Tyr Arg Arg Ile Thr Ger Ser Lys Cys Pro Lys Glu Ala Val Ile Phe Lys Thr Ile Val Ala Lys Glu Ile Grand Ala Ala Cro Lys Gln Lys Trp Val Gln Asp Ser Met Asp His Leu Asp Lys Gln Thr Gln Thr Pro Lys Thr encoded by the cDNA sequence (SEQ ID NO: 33) of:

- 1 CTAACCCAGA AACATCCAAT TCTCAAACTG AAGCTCGCAC TCTCGCCTCC
- 10 51 AGCATGAAAG TCTCTGCCGC CCTTCTGTGC CTGCTGCTCA TAGCAGCCAC
  - 101 CTTCATTCCC CAAGGGCTCG CTCAGCCAGA TGCAATCAAT GCCCCAGTCA
  - 151 CCTGCTGTTA TAACTTCACC AATAGGAAGA TCTCAGTGCA GAGGCTCGCG
  - 201 AGCTATAGAA GAATCACCAG CAGCAAGTGT CCCAAAGAAG CTGTGATCTT
  - 251 CAAGACCATT GTGGCCAAGG AGATCTGTGC TGACCCCAAG CAGAAGTGGG
- 15 301 TTCAGGATTC CATGGACCAC CTGGACAAGC AAACCCAAAC TCCGAAGACT
  - 351 TGAACACTCA CTCCACAACC CAAGAATCTG CAGCTAACTT ATTTTCCCCT
  - 401 AGCTTTCCCC AGACACCCTG TTTTATTTTA TTATAATGAA TTTTGTTTGT
  - 451 TGATGTGAAA CATTATGCCT TAAGTAATGT TAATTCTTAT TTAAGTTATT
  - 501 GATGTTTTAA GTTTATCTTT CATGGTACTA GTGTTTTTTA GATACAGAGA
  - 551 CTTGGGGAAA TTGCTTTTCC TCTTGAACCA CAGTTCTACC CCTGGGATGT
    - 601 TTTGAGGGTC TTTGCAAGAA TCATTAATAC AAAGAATTTT TTTTAACATT
    - 651 CCAATGCATT GCTAAAATAT TATTGTGGAA ATGAATATTT TGTAACTATT
    - 701 ACACCAAATA AATATATTTT TGTAC

which corresponds to the cDNA sequence and derived amino acid sequence monocyte chemoattractant protein 1 (MCP-1). See Yoshimura et al., FEBS Lett. 244, 487-493 (1989).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 34) of:

Met Lys Ala Ser Ala Ala Leu Leu Cys Leu Leu Leu Thr Ala Ala Ala Phe Ser Pro Gln Gly Leu Ala Gln Pro Val Gly Ile Asn Thr Ser Thr Thr Cys Cys Tyr Arg Phe Ile Asn Lys Lys Ile Pro Lys Gln Arg Leu Glu Ser Tyr Arg Arg Thr Thr Ser Ser His Cys Pro Arg Glu Ala Val Ile Phe Lys Thr Lys Leu Asp Lys Glu Ile Cys Ala Asp Pro Thr Gln Lys Trp Val Gln Asp Phe Met Lys His Leu Asp Lys Lys Thr Gln Thr Pro Lys Leu encoded by the cDNA sequence (SEQ ID NO: 35) of:

- 1 AGCAGAGGGG CTGAGACCAA ACCAGAAACC TCCAATTCTC ATGTGGAAGC
- 51 CCATGCCTC ACCTCCAAC ATGAAAGCCT CTGCAGCACT TCTGTGTCTG
- 101 CTGCTCACAG CAGCTGCTTT CAGCCCCCAG GGGCTTGCTC AGCCAGTTGG

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- 151 GATTAATACT TCAACTACCT GCTGCTACAG ATTTATCAAT AAGAAAATCC
- 201 CTAAGCAGAG GCTGGAGAGC TACAGAAGGA CCACCAGTAG CCACTGTCCC
- 251 CGGGAAGCTG TAATCTTCAA GACCAAACTG GACAAGGAGA TCTGTGCTGA
- 301 CCCCACACAG AAGTGGGTCC AGGACTTTAT GAAGCACCTG GACAAGAAAA
- 351 CCCAAACTCC AAAGCTTTGA ACATTCATGA CTGAACTAAA AACAAGCCAT
  - 401 GACTTGAGAA ACAAATAATT TGTATACCCT GTCCTTTCTC AGAGTGGTTC
  - 451 TGAGATTATT TTAATCTAAT TCTAAGGAAT ATGAGCTTTA TGTAATAATG
  - 501 TGAATCATGG TTTTTCTTAG TAGATTTTAA AAGTTATTAA TATTTTAATT
  - 551 TAATCTTCCA TGGATTTTGG TGGGTTTTGA ACATAAAGCC TTGGATGTAT
  - 601 ATGTCATCTC AGTGCTGTAA AAACTGTGGG ATGCTCCTCC CTTCTCTACC
  - 651 TCATGGGGGT ATTGTATAAG TCCTTGCAAG AATCAGTGCA AAGATTTGCT
  - 701 TTAATTGTTA AGATATGATG TCCCTATGGA AGCATATTGT TATTATATAA
  - 751 TTACATATTT GCATATGTAT GACTCCCAAA TTITCACATA AAATAGATTT
  - **801 TTGTAAAAA**
- which corresponds to the cDNA sequence and derived amino acid sequence monocyte chemoattractant protein 3 (MCP-3). See: Structural and Functional Identification of Two Human, Tumor-derived Monocyte Chemotactic Proteins (MCP-2 and MCP-3) Belonging to the Chemokine Family. Jo Van Damme, Paul Proost, Jean-Pierre Lenaerts, and Ghislain Opdenakker. J. Exp. Med. 176: 59-65, 1992.
- In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 36) of:

Met Lys Val Ser Ala Ala Arg Leu Ala Val Ile Leu Ile Ala Thr Ala Leu Cys Ala Pro Ala Ser Ala Ser Pro Tyr Ser Ser Asp Thr Thr Pro Cys Cys Phe Ala Tyr Ile Ala Arg Pro Leu Pro Arg Ala His Ile Lys Glu Tyr Phe Tyr Thr Ser Gly Lys Cys Ser Asn Pro Ala Val Val Phe Val Thr Arg Lys Asn Arg Gin Val Cys Ala Asn Pro Glu Lys Lys Trp Val Arg Glu Tyr Ile Asn Ser Leu Glu Met Ser

encoded by the cDNA sequence (SEQ ID NO: 37) of:

- 1 CCTCCGACAG CCTCTCCACA GGTACCATGA AGGTCTCCGC GGCACGCCTC
- 51 GCTGTCATCC TCATTGCTAC TGCCCTCTGC GCTCCTGCAT CTGCCTCCCC
- 101 ATATTCCTCG GACACCACAC CCTGCTGCTT TGCCTACATT GCCCGCCCAC
- 151 TGCCCCGTGC CCACATCAAG GAGTATTTCT ACACCAGTGG CAAGTGCTCC
- 201 AACCCAGCAG TCGTCTTTGT CACCCGAAAG AACCGCCAAG TGTGTGCCAA
- 251 CCCAGAGAAG AAATGGGTTC GGGAGTACAT CAACTCTTTG GAGATGAGCT
- 301 AGGATGGAGA GTCCTTGAAC CTGAACTTAC ACAAATTTGC CTGTTTCTGC
- 35 351 TTGCTCTTGT CCTAGCTTGG GAGGCTTCCC CTCACTATCC TACCCCACCC
  - 401 GCTCCTTGAA GGGCCCAGAT TCTGACCACG ACGAGCAGCA GTTACAAAAA

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- 451 CCTTCCCCAG GCTGGACGTG GTGGCTCAGC CTTGTAATCC CAGCACTTTG
- 501 GGAGGCCAAG GTGGGTGGAT CACTTGAGGT CAGGAGTTCG AGACAGCCTG
- 551 GCCAACATGA TGAAACCCCA TGTGTACTAA AAATACAAAA AATTAGCCGG
- 601 GCGTGGTAGC GGGCGCCTGT AGTCCCAGCT ACTCGGGAGG CTGAGGCAGG
- 651 AGAATGGCGT GAACCCGGGA GCGGAGCTTG CAGTGAGCCG AGATCGCGCC

  - 751 AAAAAAAAA AAAAAATACA AAAATTAGCC GCGTGGTGGC CCACGCCTGT
  - 801 AATCCCAGCT ACTCGGGAGG CTAAGGCAGG AAAATTGTTT GAACCCAGGA
  - 851 GGTGGAGGCT GCAGTGAGCT GAGATTGTGC CACTTCACTC CAGCCTGGGT
- 10 901 GACAAAGTGA GACTCCGTCA CAACAACAAC AACAAAAAGC TTCCCCAACT
  - 951 AAAGCCTAGA AGAGCTTCTG AGGCGCTGCT TTGTCAAAAG GAAGTCTCTA
  - 1001 GGTTCTGAGC TCTGGCTTTG CCTTGGCTTT GCAAGGGCTC TGTGACAAGG
  - 1051 AAGGAAGTCA GCATGCCTCT AGAGGCAAGG AAGGGAGGAA CACTGCACTC
  - 1101 TTAAGCTTCC GCCGTCTCAA CCCCTCACAG GAGCTTACTG GCAAACATGA
- 15 1151 AAAATCGGGG

which corresponds to the cDNA sequence and derived amino acid sequence Human T cell-specific protein (RANTES). See Schall et al., J. Immunol. 141, 1018-1025 (1988).

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 38) of:

Met Arg Ile Ser Ala Thr Leu Leu Cys Leu Leu Ile Ala Ala Ala Ala Phe Ser Ile Gln Val
Trp Ala Gln Pro Asp Gly Pro Asn Ala Ser Thr Cys Cys Tyr Val Lys Lys Gln Lys Ile
Pro Lys Arg Asn Leu Lys Ser Tyr Arg Arg Ile Thr Ser Ser Arg Cys Pro Trp Glu Ala
Val Ile Phe Lys Thr Lys Lys Gly Met Glu Val Cys Arg Glu Ala His Gln Lys Trp Val
Glu Glu Ala Ile Ala Tyr Leu Asp Met Lys Thr Pro Thr Pro Lys Pro

- 25 encoded by the cDNA sequence (SEQ ID NO: 39) of:
  - 1 ACTGAAGCCA GCTCTCTCAC TCTCTTTCTC CACCATGAGG ATCTCTGCCA
  - 51 CGCTTCTGTG CCTGCTGCTC ATAGCCGCTG CTTTCAGCAT CCAAGTGTGG
  - 101 GCCCAACCAG ATGGGCCCAA TGCATCCACA TGCTGCTATG TCAAGAAACA
  - 151 AAAGATCCCC AAGAGGAATC TCAAGAGCTA CAGAAGGATC ACCAGTAGTC
  - 201 GGTGTCCCTG GGAAGCTGTT ATCTTCAAGA CAAAGAAGGG CATGGAAGTC
    - 251 TGTCGTGAAG CCCATCAGAA GTGGGTCGAG GAGGCTATAG CATACTTAGA
    - 301 CATGAAAACC CCAACTCCAA AGCCTTGAAG AAATGTGCCT GAACAGAAAC
    - 351 CAACCTAGGA GCCAAGAAGC AAAAATTCCT CACCGCTGTT CTTTCTGAGA
    - 401 ACTGTTGATG AAATGTGTTG ATCACGGTCC TAAGGGATAG GAGCTGTCTG
- 35 451 TAGGAATGTG AAACAGTCAC GCCTAAGGAA TGGTCTTTAA GTTATTAATA
  - 501 TITTTATTTA ATTAGCCATG TACTTTGGTG TGATTTGAAT GTAAAGCTCT

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551 GGAGACCTCA TGTCACTTTA ACATTGTGTT AGCTGCAGAA TTC which corresponds to the cDNA sequence and derived amino acid sequence human fic (growth factor-activated gene). See Heinrich et al., Molecular and Cellular Biology 13: 2020-2030, 1993.

In another aspect, the present invention provides a heparanase having the amino acid sequence (SEQ ID NO: 40) of:

Asp Ser Val Ser Ile Phe Ile Thr Cys Cys Phe Asn Val Ile Asn Arg Lys Ile Pro Ile Gln Arg Leu Glu Ser Tyr Thr Arg Ile Thr Asn Ile Gln Cys Pro Lys Glu Ala Val Ile Phe Lys Thr Gly Lys Glu Val Cys Ala Asp Pro Lys Glu Arg Trp Val Arg Asp Ser Met Lys His Lys Asp Gln Ile Phe Gln Asn Leu Lys Pro

which corresponds to the cDNA sequence and derived amino acid sequence monocyte chemoattractant protein 2 (MCP-2). See VanDamme et al., J. Exp. Med. 176: 59 - 65, 1992.

The purified heparanase of the present invention, allows for the convenient selection of compounds having anti-heparanase activity (AHA compounds), i.e. inhibitors of heparanase 15 activity (IHA), by measuring inhibition of heparanase activity. Inhibition of heparanase activity can be measured utilizing in vivo radiolabeled heparan sulfate/heparin. This ligand is radiolabeled to high specific activity by intraperitoneal injection of 0.5mCi of S-35 sulfate into C57 mice bearing a 1-2 cm basement membrane tumor (EHS; Engelbreth, Holm, Swarm tumor). The tumor is harvested after 16 hours and the heparan sulfate proteoglycan extracted in 4 volumes of 6M urea, 20mM Tris pH 6.8, protease inhibitors, 0.15M NaCl and 0.5% triton X-100. The urea extract is chromatographed on an anion exchange column and the proteoglycan is eluted in a linear gradient of NaCl. The radiolabeled proteoglycan is exchanged into a solution of 4.0M guanidine-HCl, 20mM Tris pH 7.4 and applied to a size exclusion column. The proteoglycan peak is pooled and exchanged into 0.15mM NaCl and 20mM Tris pH7.4.

Purified, radiolabeled proteoglycan is coupled to commercially available agarose support. A quantitative assay of heparanase activity is constructed with the radiolabeled ligand in a multiwell format. Briefly, known quantities of recombinant heparanase are added to a multi-well plate containing equal amounts of radiolabeled ligand in each well. Enzyme-ligand interaction proceeds overnight and the ligand-agarose complex is recovered by centrifugation. Radioactivity 30 in the liquid phase is determined by scintillation counting and is the measure of enzyme activity. Potential enzyme inhibitors can be evaluated by adding the compound to the solution phase or alternatively adding the assay components to multi-well plates containing preweighed amounts of test compound.

In addition, the purified heparanase of the subject invention can be used for therapeutic wound healing or can be immobilized onto filters and used to degrade heparin from the blood of patients post-surgery.

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Wound treatment can be achieved by administration to an afflicted individual an effective amount of a pharmaceutical composition comprising the purified heparanase in combination with a pharmaceutically acceptable, preferably slow releasing, carrier. See. e.g. PCT/US90/04772, incorporated herein by reference.

Immobilization onto filters can be achieved by the methods well known in the art including those disclosed by Langer et al. in *Biomaterials: Inter-facial Phenomenon and Applications*, eds. Cooper et al, pp 493-509, 1982 and those described in U.S. Patent No. 4.373,023, 4.863,611 and 5,211,850 (all incorporated herein by reference).

The purified heparanase of the subject invention can be prepared by the method described in procedure A or procedure B, but preferably procedure A.

#### PROCEDURE A

Reverse transcription of the mRNA from activated human leukocyte-derived cells [preferably lymphocytes, neutrophils, platelets, Jurkatt lymphoma cells, Dami cells (Greenberg et al., Blood 72:1968-1977, (1988)] is used to prepare the cDNA for the desired heparanase enzyme (preferably SEQ. ID. NO: 1; optionally SEQ. ID. NO: 3, SEQ. ID. NO: 5, SEQ. ID. NO: 7; SEQ. ID. NO: 13, SEQ. ID. NO: 15, SEQ. ID. NO: 17, SEQ. ID. NO: 19, SEQ. ID. NO: 21, SEQ. ID. NO: 23, SEQ. ID. NO: 25, SEQ. ID. NO: 27, SEQ. ID. NO: 29; SEQ. ID. NO: 31, SEQ. ID. NO: 33 or SEQ. ID. NO: 35), employing standard PCR cloning techniques (described in Sambrook et al., in: Molecular Cloning, A Laboratory Manual. Second Edition, 1989. Cold Spring Harbor Press). The cDNA encoding the heparanase enzyme is cloned into Xba1/BamH1 sites in the commercially available baculovirus vector pVL 1392 (Pharmingen; San Diego, CA). High titer infectious virus is selected for use in infecting sf9 insect cells (Luckow and Summers, BiolTechnology. 6,47 1988). Serum-free medium conditioned by infected sf9 cells is collected after 72 hours. This media is the starting material for purification of recombinant heparanase. Serum-free conditioned media is adjusted to contain 20mM Sodium Acetate, pH 5.0, 0.15M NaCl, 1mM reduced glutathione (GSH), 1mM dithiothreitol (DTT) and 10mM beta-octylglucoside. Medium is applied to a column of cation-exchange resin (Pharmacia) and eluted from the column in a linear gradient of NaCl. Fractions containing heparanase are pooled and diluted to a final salt concentration of 0.15M NaCl. To this solution is added 20mM Tris and the pH adjusted to 7.0. The solution is applied to a column of heparin-Sepharose (Pharmacia) and eluted with a linear salt gradient buffered to pH 5.0 with 20mM Sodium Acetate. Heparanase is concentrated to 0.5mg/ml in an Amicon concentrator fitted with a YM-2 membrane and stored at -80 degrees. For optimal activity (greater than 50 units heparanase activity per ug protein) incubation in the presence of transglutaminase, under reducing conditions, in accordance with the procedure in Example 2, Part C.

### PROCEDURE B

This procedure describes the purification to homogeneity of heparanase (SEQ. ID. NO: 1) from human blood cells or cell lines (such as platelets) under reducing conditions which allow for the occurrence of post-translational modifications that increase the specific activity of heparanase and make it suitable for use in the above described screening assay. The cells are treated with a suitable activator (such as, but not limited to, thrombin or histamine) which allows for the release of enzymes and cytokines from the cell. Reducing agents are added to the supernatant from the activated cells. Suitable reducing agents include dithiothreitol (DTT), dithioerythritol (DTE), reduced glutathione (GSH), and β-mercaptoethanol. The reduced, activated supernatant is chromatographed on a column of immobilized heparin or heparan sulfate under reducing conditions at pH 5, using a salt gradient (such as NaCl, KCl, or other salt) to elute the bound proteins. Fractions containing heparanase activity are pooled and exchanged into any buffer appropriate for the pH of 6.8 and containing 0.15 M NaCl, reducing agents, and non-ionic detergent. This is passed over any suitable anion-exchange column (bed volume of 5 ml or less). The unbound material from this column is adjusted to pH 5 with acid, and is loaded onto any suitable cation-exchange column (bed volume of 5 ml or less), equilibrated in a suitable pH 5 buffer containing 0.15 M NaCl, reducing agents, and non-ionic detergents. The bound protein is eluted from the column with a salt gradient, and the fractions containing heparanase activity are pooled and size fractionated to below 30,000 daltons with 30 K-cut-off membranes. The protein below 30,000 daltons is concentrated by either heparin-sepharose chromatography or by centrifugation through 5 K-cut-off membranes.

The present invention is seen more fully by the examples set forth below.

Example 1: Use of Heparanase as a screen for AHA compounds.

- 1. Heparan sulfate, metabolically labeled (S-35) to a high-specific activity- as described above for the EHS tumor, prepared by papain digestion of chromatographically purified heparan sulfate proteoglycan is coupled to cyanogen bromide activated Sepharose-6B (Pharmacia) according to manufacturer's instructions.
  - 2. <sup>35</sup>S-Heparan sulfate-Sepharose 6B is resuspended in: 0.15 M NaCl, 0.03% human serum albumin, 10 μM MgCl<sub>2</sub>, 10 μM CaCl<sub>2</sub>, antiproteolytic agents (1 μg/ml leupeptin, 2 μg/ml antipain, 10 μg/ml benzamidine, 10 units/ml aprotinin, 1 μg/ml chymostatin, and 1 μg/ml pepstatin), and 0.05 M Na acetate, pH 5.6 and 5,000 cpm, in a total volume of 200 μl, are aliquoted into each well of a 96 well plate. To each well is added 5 units of activated heparanase and the digestion allowed to proceed overnight at 37 degrees.
- Separation of digested product is accomplished by centrifugation of the 96 well
   plate. The supernatant, containing cleaved heparan sulfate is decanted and quantitated by scintillation counting.

- Inhibitors of heparanase activity can be introduced into the liquid-phase of the assay.
- 5. A potential inhibitor of heparanase activity would be identified by its ability to reduce the amount of radiolabeled heparan sulfate released into the supernatant by 50% at a concentration of 1 µM or less.

Example 2: The preparation of heparanase under reducing conditions as outlined in Procedure B.

#### Part A:

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Platelet-rich plasma (10<sup>9</sup> platelets/ml; 1800 ml) is obtained from healthy, informed volunteers by plasmapheresis. The plasma is removed from the platelets by centrifugation (Heldin, et al., Exp. Cell Res. 109: 429-437, 1977). Platelets suspended in phosphate buffered saline (PBS; 0.1 original volume) are then stimulated with 1 U/ml thrombin for 5 min at 37°C. This concentration of thrombin was reported to release 100% of the heparanase activity from platelets (Oldberg, et al., Blochemistry 19: 5755-5762, 1980). Following activation, the thrombin is inactivated by the addition of 100 mM phenylmethylsulfonylfluoride (PMSF), and the platelets are centrifuged at 2000 x g for 30 min at 4°C. The supernatant is stored at -80°C until used for the chromatographic purification of heparanase (Part B).

Part B: Chromatographic purification of heparanase.

- 1. Heparin-Sepharose Chromatography. Activated platelet supernatants are pooled and adjusted to contain 1 mM GSH and 1 mM DTT. This pool is loaded (2.5 ml/min) onto a column of heparin-sepharose (2.6 x 7.5 cm, 40 ml) equilibrated in 1 mM GSH, 1 mM DTT, 150 mM NaCl, 10 mM NaPO<sub>4</sub>, pH 7.4. After loading the sample, the column is washed with 200 ml of 0.15 M NaCl, 1 mM GSH, 1 mM DTT, 10 mM Na acetate, pH 5, followed by 60 ml of 0.35 M NaCl, 1 mM DTT, 1 mM GSH, 10 mM Na acetate, pH 5. The column is then eluted with a 160 ml linear gradient between 0.35 M NaCl and 1.5 M NaCl in the same buffer. Aliquots of each fraction are used for determination of heparanase activity by the "Purification Assay" described later.
- 2. Anion-exchange chromatography (For example, DEAE-Sephacel, Pharmacia). The 0.9 M 1.15 M NaCl fractions from the heparin-sepharose column are concentrated using a stirred cell fitted with a PM-10 membrane, and the buffer is exchanged to 0.15 M NaCl, 1 mM DTT, 1 mM GSH, 10 mM β-octylglucoside, 10 mM sodium phosphate, pH 6.8 (8 ml). This sample is loaded onto a 5 ml column of DEAE-Sephacel equilibrated in the same buffer. After loading, the column is washed to baseline absorbance (280 nm) with the equilibration buffer. The flow-through and wash with equilibration buffer are collected as one pool. The column is then eluted with 10 ml of 0.15 M NaCl, 10 mM β-octylglucoside, 1 mM GSH, 1 mM DTT, 10 mM Na acetate, pH 5, followed by 10 ml of 1.5 M NaCl, 10 mM β-octylglucoside, 1 mM GSH, 1 mM

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DTT, 10 mM Na acetate, pH 5. Aliquots of each pool are used for determination of heparanase activity by the "Purification Assay".

- 3. Cation Exchange. The unbound sample from the DEAE-Sephacel column is adjusted to pH 5 with glacial acetic acid and loaded onto a cation exchange column (Poros HS/F, 4.6 mm x 50 mm; PerSeptive Biosystems), pre-equilibrated with 0.15 M NaCl, 1 mM DTT, 1 mM GSH, 10 mM β-octylglucoside, 10 mM Na acetate, pH 5. The flow rate is 3.0 ml/min and 3 ml fractions are collected. After washing the column with 35 ml of equilibration buffer, the column is developed with a 55 ml linear gradient between 0.15 M and 1.5 M NaCl in the same buffer. 10 μl aliquots of the fractions are used for determination of heparanase activity by the "Purification Assay".
- 4. Size fractionation to < 30 kD and concentration on immobilized heparin (Hi-trap heparin-sepharose, Pharmacia). The activity from the Poros HS/F column is size fractionated by centrifuging through 30,000 molecular weight cut-off filters (Millipore ultrafree-MC 30,000 NMWL filter units). The < 30 kD pool is diluted to contain 0.15 M NaCl, and is loaded onto a 1 ml Hi-trap heparin column, pre-equilibrated with 0.15 M NaCl, 1 mM DTT, 1 mM GSH, 10 mM Na acetate, pH 5. The column is eluted with 1.2 M NaCl in the same buffer and the single eluted peak contains the heparanase activity.

Part C: Properties of the purified heparanase.

The final yield of heparanase protein from 1850 ml platelet-rich plasma was 2.7 mg. Protein concentration was determined by the method of Lowry (*J. Biol. Chem.* 193: 265-275, 1951), or if more precise determinations were required, by amino acid analysis on an amino acid analyzer (Beckman 6300). The overall recovery of activity was 8%, with a 4150-fold purification. The preparation was judged to be homogeneous by the presence of a single band of 9000 daltons on an 18% silver-stained SDS-polyacrylamide gel, run according to the method of Laemmli (*Nature* 227: 680-685, 1970).

The pH optimum of the purified heparanase was determined by conducting the "Purification assay" activity between pH 3.5 and 8.0, using a citrate buffer (pH 3.5 - 6.0), citrate-phosphate buffer (pH 6.5 - 7.0), and phosphate buffer (pH 7.5 - 8.). Heparanase was active between pH 5.0 and 8.0, with the optimum pH at 5.8.

N-terminal amino acid sequencing of heparanase produced by this procedure was performed using a gas/liquid phase Protein Sequencer (Applied Biosystems Inc. Model 470). Phenylthiodantoin amino acids were resolved and quantitated by an on-line HPLC system (Model 120, Applied Biosystems Inc.) with data analysis on a Nelson Analytical System. N-terminal amino acid sequences of the heparanase produced in this example were 85 %

35 SEQ. ID. NO: 9 (namely:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala
1 5 10 15
Glu Leu Arg).

which is identical to CTAP-III, and 15% SEQ. ID. NO: 10 (namely:

5 Ser Ser Thr Lys Gly Gin Thr Lys Arg Asn Leu Ala Lys Gly Lys Glu), 1 5 10 15

which is the precursor form, platelet basic protein. Interestingly, the N-terminal sequence of commercial β-thromboglobulin (namely, Calbiochem (Cat. # 605165), Celsus Laboratories (Cat. # 41705), and Haematologic Technologies (Cat. # HBTG-0210), which has low levels of

10 heparanase activity, was 100% SEQ. ID. NO: 11 (namely:

Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Tyr Ala Glu),

1 10 15

indicating that the commercial preparatio: is actually CTAP-III and not β-thromboglobulin.

Chromatofocusing of the heparanase produced by this procedure results in two peaks of differing isoelectric points. To perform the chromatofocusing, heparanase is dissolved in 0.025 M imidazole, pH 7.3. The sample is loaded onto a 0.5 x 20 cm column of Polybuffer Exchanger 94 (Pharmacia), equilibrated with 0.025 M imidazole, pH 7.3. Immediately after sample loading, Polybuffer 74 (Pharmacia; 1:8, pH 4) is pumped onto the column at 0.5 ml/min. 2 ml fractions are collected, and the pH of each fraction is determined by a narrow range (pH 4-20 7) pH paper. Aliquots of each fraction are used to determine heparanase activity by the "Purification Assay." All of the activity is associated with an absorbance (280 nm) peak that eluted at pH 4.8 to 5.1, representing approximately 10% of total protein, while 90% of the protein is eluted at pH 7.3 and is inactive. Aliquots of each protein peak are separated from the ampholytes by C<sub>4</sub> reverse phase chromatography. The peak that eluted from the chromatofocusing column at pH 7.3 has N-terminal sequences for platelet basic protein and the processed form, CTAP-III. The peak that is eluted from the chromatofocusing column at pH 4.8 - 5.1 also contains the sequences of platelet basic protein and the processed form, CTAP-III. All of the platelet basic protein processed forms have pl's that are calculated and reported to be greater than 7.6. Thus, the heparanase activity resides in the platelet basic protein and/or the processed

Heparanase obtained after chromatofocusing exhibited a specific activity of 80 unitsing protein, using the "Purification Assay." This represents a 1000-fold increase in the specific activity compared to the commercial protein (β-thromboglobulin; 0.075 units/μg protein).

form, CTAP-III that is modified such that the pI is lowered to 4.8 - 5.1.

The modification that may be responsible for the lower isoelectric point of active

begin to heparanase is ADP-ribosylation. ADP-ribosylation (Adenine diphosphate-ribosylation) is a posttranslational modification of proteins or DNA in which the ADP-ribose group of NAD

(Nicotinamide adenine dinucleotide) is enzymatically transferred to proteins or DNA. Since this

modification adds two negatively charged phosphate groups to a molecule, it would result in a lower isoelectric point. Activated platelet supernatants were incubated in 1 mM DTT, 2 mM MgCl<sub>2</sub>, 100 mM HEPES, pH 7.4, and 0.5 μM [<sup>32</sup>P]NAD (Specific activity = 1000 Ci/mmol). The labeled proteins were separated by SDS-polyacrylamide gel electrophoresis on an 18% gel, transferred to PVDF (polyvinylidene difluoride) membrane, and exposed to X-ray film. The autoradiogram demonstrates the incorporation of [32P] into a protein of 8000 daltons. The PVDF membrane was immunoblotted with the anti-Peptide C antisera (1:1500 in PBS containing 5% dry milk, 0.05% Tween-20, 0.15 M NaCl, 20 mM Tris, pH 7.4, 2 hours at room temperature, followed by incubation with peroxidase-labeled goat anti-chicken IgG (1:500 in above buffer, 1 hour room temperature), and reacted with a peroxidase substrate. The immunoblot revealed that the 8000 dalton that was labeled with [32P] was CTAP-III/heparanase. The addition of 200 µM sodium nitroprusside, a spontaneous releaser of nitric oxide, to the ADP-ribosylation reaction resulted in 5-fold more incorporation of [32P] label into CTAP-III/heparanase, suggesting that this modification can be regulated in vivo by nitric oxide. 15 Finally, in an analgous manner to that of glyceraldehyde-3-phosphate dehydrogenase, another platelet ADP-ribosylated glycolytic enzyme (Zhang and Snyder, Proc. Natl. Acad. Sci. USA 89: 9382-9385), it was determined that CTAP-III/heparanase has an auto-ADP-ribosylation activity, since the [32P]-ADP-ribosylation of CTAP-III/heparanase occurs in reactions where the only protein present is commercial CTAP-III or purified heparanase. Other chemokine family members tested, which includes IP-1-, IL-8, gro-a, and MCAF, also have auto-ADP-ribosylation activity.

It is contemplated that the high specific activity of CTAP-III/heparanase is a consequence of ADP-ribosylation of the enzyme in the presence of nitric oxide. It is further contemplated that the action of transglutaminase on the ADP-ribosylated enzyme will lead to further increase in the specific activity.

An amino acid composition of the heparanase produced in Example 2 gave the expected amino acid composition for CTAP-III and N-terminal sequencing revealed sequences for platelet basic protein and the processed form, CTAP-III, confirming that the heparanase activity is contained in this set of processed proteins and is not due to a minor contaminant. The presence of heparanase activity in three commercial sources of  $\beta$ -thromboglobulin also confirms this conclusion. In addition, polyclonal antibodies to  $\beta$ -thromboglobulin were found to precipitate 30 - 70% of the heparanase activity in three separate experiments, providing additional confirmation.

The activation of heparanase with transglutaminase (prepared in accordance with Example 2, Part B) results in a substantial (about 13-fold) increase in the specific activity of the enzyme. The heparanase (2 ul at 56 nM) obtained by Example 2, Part B is treated with either

transglutaminase from guinea pig liver (4 mU; Sigma) or with Factor XIII (1 µg; Celsus Laboratories, Inc.), the blood coagulation factor that is activated by treatment with 5 units of thrombin at 37 degrees for 30 minutes. Heparanase is activated by incubation of either 2mU liver transglutaminase or 5 units of activated Factor XIII in the presence of 0.1M NaAcetate buffer at pH 6.0 containing 1mM reduced glutathione and 1mM CaCl for 35 minutes at 37 degrees. Treatment of heparanase with either type of transglutaminase results in a substantial increase in the specific activity of the heparanase.

The high degree of sequence identity between CTAP-III and Interleukin-8, a CXC chemokine family member, assures that an essentially identical folding pattern will be shared by the two proteins. Since the 3-dimensional structure of Interleukin-8 is known (Clore, et al., Biochemistry 29: 1689-1696,1990; Baldwin, et al., J. Biol. Chem. 265: 6851-6853), one can model the same for CTAP-III. Such a model can serve to direct research into rationally designed IHA and to help explain the action of transglutaminase in activating the CTAP-III.

# Part D: Purification Assay for Heparanase Activity

15 Heparanase activity from platelets or column fractions is detected by its ability to digest the  $\geq$  70 kD <sup>35</sup>S-HSPG to produce lower molecular weight products. Each digest contains 10  $\mu$ l sample, <sup>35</sup>S-HSPG (2000 cpm), 0.15 M NaCl, 0.03% human serum albumin, 10 µM MgCl<sub>2</sub>, 10 μM CaCl<sub>2</sub>, antiproteolytic agents (1 μg/ml leupeptin, 2 μg/ml antipain, 10 μg/ml benzamidine, 10 units/ml aprotinin, 1 µg/ml chymostatin, and 1 µg/ml pepstatin), and 0.05 M Na acetate, pH 5.6 in a total volume of 300 µl. Digests are carried out for 3 to 21 h. The presence of lower molecular weight radiolabeled products is detected by centrifugation through 30,000 MW-cutoff filters. The digests containing 2000 cpm of <sup>35</sup>S-HSPG (> 70 K) are centrifuged through 30,000 molecular weight cut-off filters (Millipore Ultrafree-MC 30,000 NMWL filter units). 35S-HSPG degradation is evident by the presence of radioactivity in the filtrate that passed through the 30 25 K membrane; this heparanase activity is expressed as the % of total cpm < 30 K for a given digest. Analysis of heparan sulfate degradation by this method is quick and reproducible. 1 unit of heparanase activity is defined as 1% cpm < 30 K per h. For pH optimum determination, the 0.1 M Na acetate buffer is replaced by 50 mM citrate, citrate-phosphate, or phosphate buffer at varying pH's. For samples from chromatographic steps performed under reducing conditions (1 mM GSH, 1 mM DTT), the concentration of a thiol oxidant (diamide) needed for optimum activity is determined. This concentration (100 µM diamide) is added to all assay tubes when reduced samples are assayed.

# Preparation of <sup>35</sup>S-HSPG (>70 K) for use in the "Purification Assay."

<sup>35</sup>S-HSPG (>70 K) is prepared from mice bearing a basement membrane tumor that overproduces HSPG (EHS tumor), using modifications of the method of Ledbetter, et. al., 1987. Briefly, the radiolabeled HSPG was prepared by injecting C57BL mice bearing the EHS tumor

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(Orkin, et.al., 1977) with sodium [35S]sulfate (0.5 mCi/mouse) 18 h before harvesting the tumor. The HSPG is extracted from the weighed tumor with 6 volumes (w/v) of Buffer A (3.4 M NaCl, 0.1 M 6-aminohexanoic acid, 0.04 M EDTA, 0.008 M N-ethylmaleimide, 0.002 M PMSF, and 0.05 M Tris-HCl, pH 6.8), by homogenization with a Polytron for 30 s, followed by stirring at 4°C for 1 h. Insoluble material is collected by centrifugation (12,000 x g for 10 min), and the supernatant is discarded. The insoluble residue is reextracted with 2 volumes (original tumor weight) of Buffer A for 30 min with stirring at 4°C. Insoluble material is again collected by centrifugation, and the supernatant fraction is discarded. The insoluble material is then suspended in 6 volumes of Buffer B (6 M urea, 0.1 M 6-aminohexanoic acid, 0.04 M ethylenediaminetetraacetic acid (EDTA), 0.002 M PMSF, and 0.05 M Tris-HCl, pH 6.8), homogenized with an electric homogenizer (Polytron) for 30 s, and stirred for 2 h at 4°C. The mixture is centrifuged to remove insoluble material, and the supernatant is retained. The insoluble material is reextracted with 2 volumes of Buffer B. The mixture is centrifuged, and the supernatant is combined with the previous supernatant.

35S-HSPG is isolated from the Buffer B supernatant by sequential chromatography on anion exchange and gel filtration columns. The Buffer B supernatant is dialyzed overnight against 10 volumes of 6 M urea, 0.15 M NaCl, 0.05 M Tris-HCl, pH 6.8, and is adjusted to contain 0.5% non-ionic detergent (Triton X-100). This supernatant (from 11 g tumor) is chromatographed on a 30 ml column of anion exchange resin (DEAE-Sephacel) equilibrated with 6 M urea, 0.15 M NaCl, 0.05% Triton X-100, 0.05 M Tris-HCl, pH 6.8. After loading the supernatant and washing with the equilibration buffer, the column is developed with a 250 ml linear gradient between 0.15 M NaCl and 1.15 M NaCl (Flow = 2.0 ml/min). Fractions are sampled for radioactivity, and those containing the <sup>35</sup>SO<sub>4</sub> label that elutes from the DEAE-Sephacel between 0.4 M and 0.8 M NaCl are pooled. The proteoglycan is precipitated by the addition of 4 volumes of 100% EtOH at -20°C overnight. The precipitate is collected by centrifugation and is solubilized in 1 ml of Buffer C (4 M Gu-HCl, 20 mM Tris-HCl, pH 7.2). This solubilized pellet is used for chromatography on a calibrated gel filtration column (1.0  $\times$  50 cm column of Superose 6; Pharmacia) equilibrated in Buffer C (Flow = 0.5 ml/min). Fractions are sampled for radioactivity, and those containing the 35SO4 label that elutes with a molecular weight ≥ 70 kD were pooled. The proteoglycan is precipitated with 100% EtOH as described above. The pellet is dissolved in 3 ml PBS, and dialyzed against 3 x 100 volumes of PBS. Each preparation of <sup>35</sup>S-HSPG is confirmed to be ≥ 98% heparan sulfate by susceptibility to low pH nitrous acid degradation (Shiveley and Conrad, Biochemistry 15: 3932-3942, 1976). Example 3: Preparation of cDNA encoding Heparanase.

Media is removed from cultured HEL (HEL 92.1.7; Human erythroleukemia; ATCC No. TIB 180) cells stimulated with 10nM phorbol 12-myristate 13-acetate (Sigma Chemical Co., St.

Louis, MO) and the cells scraped from the dish and pelleted by centrifugation. The pellet is extracted with 200ul of TRI reagent (Molecular Research Center Inc. Cincinnati, OH) and the total cellular RNA is prepared according to the manufacturer's instructions. To prepare first strand synthesis the reverse transcriptase reaction was performed with 10ul of total cellular RNA in the presence of 4ul of 5x transcriptase buffer (Bethesda Research Laboratories, Gaithersburg, MD), 1ul 0.2mM DTT, 4ul random hexanucleotides (Amersham Corp. Arlington Heights, ILL), and 1ul 10mM dNTP (BRL). This solution is heated to 95 degrees C for 5 minutes and then placed on ice. To this is added 1ul RNAsin and 1ul reverse transcriptase (M-MLV), (Promega, Madison WI). This is incubated at 37 degrees for 60 minutes and then placed on ice. The polymerase chain reaction is carried out as follows. To 3ul of the first strand (above) is added 1ul of each Primer (see below), 77ul of water 10ul 10x PCR buffer (Perkin Elmer Cetus, Norwalk CT) and 2ul each dNTP. This solution is heat denatured at 95 degrees C and 1ul Amplitaq DNA polymerase (Perkin Elmer Cetus) is added. Hybridization temperature begins at 72 degrees and is lowered by one degree per cycle until reaching 55 degrees. Each hybridization step is followed with a constant elongation temperature of 72 degrees. Upon completion the solution is left at 0 degrees until storage at -20 degrees. The products of the PCR reaction are electrophoresed on 3% NuSieve, 1% agarose gels and bands of expected size are excised and purified by standard procedures. Primers:

Platelet Basic Protein: TGG ACT AGT ATG TCC TCC ACC AAA GGA CAA ACT AA 20 CTAP III: TGG ACT AGT ATG AAC TTG GCG AAA GAG GA B-thrombglobulin: TGG ACT AGT ATG GGC AAA GAG GAA AGT CTA GAC AG NAP-2: TGG ACT AGT ATG GAA CTC CGC TGC ATG TGT ATA AA Example 4: Preparation of cDNA encoding Heparanase.

Media is removed from cultured leukocyte-derived cells [e.g., lymphocytes, neutrophils, platelets, Jurkatt lymphoma cells, Dami cells (Greenberg et al., Blood 72:1968-1977, (1988)], stimulated with Concanavalin A or phorbol 12-myristate 13 acetate (Sigma Chemical Co., St. Louis, MO) and the cells scraped from the dish and pelleted by centrifugation. The pellet is extracted with 200ul of TRI reagent (Molecular Research Center Inc. Cincinnati, OH) and the 30 total cellular RNA is prepared according to the manufacturer's instructions. To prepare first strand synthesis the reverse transcriptase reaction was performed with 10ul of total cellular RNA in the presence of 4ul of 5x transcriptase buffer (Bethesda Research Laboratories, Gaithersburg, MD), 1ul 0.2mM DTT, 4ul random hexanucleotides (Amersham Corp. Arlington Heights, ILL), and 1ul 10mM dNTP (BRL). This solution is heated to 95 degrees C for 5 minutes and then placed on ice. To this is added 1ul RNAsin and 1ul reverse transcriptase (M-MLV), (Promega,

WO 95/04158

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Madison WI). This is incubated at 37 degrees for 60 minutes and then placed on ice. The polymerase chain reaction is carried out as follows. To 3ul of the first strand (above) is added 1ul of each Primer (see below), 77ul of water 10ul 10x PCR buffer (Perkin Elmer Cetus, Norwalk CT) and 2ul each dNTP. This solution is heat denatured at 95 degrees C and 1ul Amplitaq DNA polymerase (Perkin Elmer Cetus) is added. Hybridization temperature begins at 72 degrees and is lowered by one degree per cycle until reaching 55 degrees. Each hybridization step is followed with a constant elongation temperature of 72 degrees. Upon completion the solution is left at 0 degrees until storage at -20 degrees. The products of the PCR reaction are electrophoresed on 3% NuSieve, 1% agarose gels and bands of expected size are excised and purified by standard procedures.

Platelet Basic Protein: TGG ACT AGT ATG TCC TCC ACC AAA GGA CAA ACT AA
CTAP III: TGG ACT AGT ATG AAC TTG GCG AAA GAG GA

B-thrombglobulin: TGG ACT AGT ATG GGC AAA GAG GAA AGT CTA GAC AG

NAP-2: TGG ACT AGT ATG GAA CTC CGC TGC ATG TGT ATA AA

All temperatures expressed throughout the subject specification are in degrees Centigrade.

The cDNA encoding heparanase is preferably cloned into a vector designed for expression in eukaryotic cells, rather than into a vector designed for expression in prokaryotic cells (e.g. E. coli). Eukaryotic cells are preferred for expression of genes obtained from higher eukaryotes because the signals for synthesis, processing, and secretion of these proteins are usually recognized, whereas this is often not true for prokaryotic hosts (Ausubel, et al., ed., in Short Protocols in Molecular Biology, 2nd edition, John Wiley & Sons, publishers, pg.16-49, 1992.). Eukaryotic hosts may include, but are not limited to, the following: insect cells, African green monkey kidney cells (COS cells), Chinese hamster ovary cells (CHO cells), and Murine 3T3 fibroblasts.

Experiments demonstrating that a synthetic peptide of CTAP-III/NAP-2 or antisera raised against a synthetic peptide of CTAP-III/NAP-2 inhibit the heparanase activity of CTAP-III/NAP-2 suggest that the amino acids participating in enzymatic catalysis are contained in a C-terminal region of the enzyme.

Peptide Synthesis: A C-terminal peptide contained within the sequences known for CTAP-III (SEQ ID NO: 1), Platelet Basic Protein (SEQ ID NO: 3), β-thromboglobulin (SEQ ID NO: 5), and NAP-2 (SEQ ID NO: 7), and an N-terminal peptide contained within the sequences known for CTAP-III (SEQ ID NO: 1) and Platelet Basic Protein (SEQ ID NO: 3) were synthesized according to standard procedures. The N-terminal peptide has the following sequence (SEQ ID NO: 41: Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Cys, in which the final Cys residue was added to regions of known sequence (SEO ID NOS: 1,3) for the purpose of

conjugation to a carrier protein. The C-terminal peptide has the following sequence (SEQ ID NO: 42): Cys Asn Gln Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys encoded by the cDNA sequence (SEQ ID NO: 43) of TGCAACCAAG TCGAAGTGAT AGCCACACTG AAGGATGGGA GGAAAATCTG CCTGGACCCA GATGCTCCCA GAATCAAGAA AATTGTACAG AAAAAA. These peptides (SEQ ID NOS: 41 and 42) were produced by stepwise solid phase peptide synthesis on an Applied Biosystems 430A Peptide Synthesizer. The 9fluoroenylmethyloxycarbonyl (Fmoc) group was used as the  $N^{\alpha}$  amino protecting group, and temporary side-chain protectin groups were as follows: Arg (Pmc), Asn (Trt), Asp (OtBu), Gln 10 (Trt), Glu (OtBu), His (Trt), Lys (Boc), Ser (tBu), Thr (tBu). Each residue was single coupled using a HBTU/NMP protocol and capped with acetic anhydride before the next synthesis cycle. After removal of the N-terminal Fmoc group, temporary side-chain protecting groups were removed and the peptide cleaved from the resin by treatment with 95% TFA/5% scavengers (ethyl methyl sulfide/aniscial), 2-ethanedithiol, 1:3:1) for two hours at room temperature. The crude peptides were precipitated from the cleavage solution with cold diethyl ether. The 15 precipitated peptide was collected on a sintered glass funnel, washed with diethyl ether, dissolved in dilute acetic acid, evaporated to dryness under reduced pressure, and the residue was redissolved and lyophillized from glacial acetic acid. The crude peptides were purified by preparative reverse phase chromatography on a Phenomenex C-18 column (22.5 x 250 mm) using a water/acetonitrile gradient, each phase containing 0.1% trifluoracetic acid (TFA). Clean fractions, as determined by analytical HPLC, were pooled, the acetonitrile was evaporated under reduced pressure, and the aqueous solution was lyophillized. The purified peptides were characterized by time of flight or FAB mass spectroscopy.

Further, SEQ ID NO: 42 can be produced by recombinant DNA methodology as stated in Procedure A (page 21).

Antisera Production: The synthetic peptides of CTAP-III/NAP-2 were conjugated to keyhole limpet hemocyanin utilizing a maleimide-activated carrier protein (Pierce Chemical Co. #77107). 300 μg of conjugated peptides were injected into chickens using Freunds complete adjuvant. The antisera were collected 5 weeks after initial immunization. Specific recognition by the antisera of commercial CTAP-III (2.5 μg, (Celsus Laboratories Inc., Cincinnati, Ohio; Cat #. 41705), isolated heparanase (1.5 μg), and 10 μl of the platelet supernatant used for purification was achieved by separating the proteins on a reducing 18% polyacrylamide gel (Novex), transferring to nitrocellulose, and incubating with the pre-immune or antisera (1:1500), followed by incubation with a peroxidase labeled goat anti-chicken IgG (1:500; Kierkegaard and Perry) in the presence of PBS containing 5% dry milk and 0.05% Tween-20. The pre-immune sera did not recognize 7 - 10 kD proteins in the commercial CTAP-III, isolated heparanase, or platelet

supernatants.

Inhibition of heparanase activity by the C-terminal synthetic peptide (SEQ ID NO: 42) or antisera: For experiments designed to determine whether the peptide antisera was able to inhibit heparanase activity, the pre-immune and antisera were exchanged into 0.15M NaCl, 0.01M sodium phoshate buffer, pH 7.4 (PBS) using a 100 kD cut-off membrane in order to remove low molecular weight chicken heparanase normally present in the serum. Aliquots of isolated heparanase (15 ng) were pre-incubated for 30 min with 2 µl of either pre-immune or anti-CTAP-III antisera before adding the 35S-HSPG to determine heparanase activity. In the presence of the pre-immune sera, the isolated protein had 14.3 ± 0.1 units of heparanase activity, while in the presence of the C-terminal peptide antisera, only  $0.8 \pm 0.2$  units of heparanase were 10 detected (p < 0.001; results confirmed in a second experiment). The N-terminal peptide antiserum was not able to neutralize the heparanase activity. Similar results were obtained when the ability of the synthetic peptides to neutralize heparanase activity was examined. Heparanase assays conducted with 3 nM enzyme, 47 nM <sup>35</sup>S-HSPG substrate, and varying concentrations of peptides showed that heparanase activity was only 5% of control values in the presence of 250 15 μM C-terminal peptide. By contrast, heparanase activity in the presence of 250 μM of either the N-terminal peptide or an unrelated peptide (PLALWAR) was 67% of control values. The ability of both the C-terminal peptide (SEQ ID NO: 42) or antisera from a chicken immunized with the C-terminal synthetic peptide to neutralize heparanase activity demonstrates conclusively that CTAP-III and NAP-2 possess heparanase activity, and suggests that the C-terminal region is essential for catalysis. Modeling of this domain (SEQ ID NO: 42) can be used in the identification of potent peptide-mimetic compounds capable of inhibiting this enzyme activity.

Computer assisted modeling can be accomplished using programs for automated docking of molecules within 3D databases, as described in DesJarlais, R.L., Sheridan, R.P., Seibel, G.L., Dixon, J.S., Kuntz, I.D., Venkataraghavan, R., "Using shape complementarity as an initial screen in designing ligands for a receptor binding site of known three-dimensional structure"; J. Med. Chem. 31:722-729, 1988. Also, automated de novo construction of ligands that can bind the catalytic site as described in Moon, J.B., Howe, W.J., "Computer design of bioactive molecules: a method for receptor-based de novo ligand design"; Proteins: Struct., Funct., and Genetics, 11:314-328, 1981.

PCT/US94/08207

#### SEQUENCE LISTING

```
(1) GENERAL INFORMATION:
           (i) APPLICANT: Hoogewerf, Arlene J.
Ledbetter, Steven R.
          (ii) TITLE OF INVENTION: USE OF HEPARANASE TO IDENTIFY AND ISOLATE ANTI-HEPARANASE COMPOUNDS
10
         (111) NUMBER OF SEQUENCES: 43
15
          (iv) CORRESPONDENCE ADDRESS:
                 (A) ADDRESSEE: The Upjohn Company, Intellectual Property Law (B) STREET: 301 Henrietta
                 (C) CITY: Kalamazoo
                 (D) STATE: MI
(E) COUNTRY: USA
20
                 (F) ZIP: 49001
           (V) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
                 (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS
25
                 (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
          (V1) CURRENT APPLICATION DATA:
                 (A) APPLICATION NUMBER:
30
                 (B) FILING DATE:
                 (C) CLASSIFICATION:
       (viii) ATTORNEY/AGENT INFORMATION:
                 (A) NAME: Jameson, William G. (B) REGISTRATION NUMBER: 27,199
35
                 (C) REFERENCE/DOCKET NUMBER: 4731.1 CP
          (ix) TELECOMMUNICATION INFORMATION:
                 (A) TELEPHONE: 616/385-7561
(B) TELEFAX: 616/385-6897
40
                 (C) TELEX: 224401
     (2) INFORMATION FOR SEQ ID NO:1:
45
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(C) STRANDEDNESS: single
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                 (D) TOPOLOGY: linear
          (ii) MOLECULE TYPE: peptide
55
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           Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp
50 60
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15	$\cdot$	
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50	Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile Ala Thr Leu 50 55 60	•
	Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys 65 70 75 80	
55	Lys Ile Val Gln Lys Lys Leu Ala Gly Asp Glu Ser Ala Asp 85 90	
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65	(b) 1010001, xx	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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25	1 5 10 15	
20	Met Cys Ile Lys Thr Thr Ser Gly Ile His Pro Lys Asn Ile Gln Ser 20 25	
	Leu Glu Val Ile Gly Lys Gly Thr His Cys Asn Gln Val Glu Val Ile	
30	35 40 45	
	Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp Pro Asp Ala Pro 50 60	
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	Asp	
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	GAT	243
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0	Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile Cys Leu Asp 35 40 45									
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40 45	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 19 amino acids  (B) TYPE: amino acid  (D) TOPOLOGY: linear									
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		1		Ser		5					10					15		
35		Leu	Gly	Leu	Leu 20	Leu	Leu	Pro	Leu	Val 25	Val	Ala	Phe	Ala	Ser 30	Ala	Glu	
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60				) ST					16							-		
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PCT/US94/08207

WO 95/04158

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65	CCTGTTAATC CAAGGTCTTT AGAAAAACTT GAAATTATTC CTGCAAGCCA ATTTTGTCCA	240
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	TCGAAGGCCA TCAAGAATTT ACTGAAAGCA GTTAGCAAGG AAATGTCTAA AAGATCTCCT	360

PCT/US94/08207

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30	Arg Val Ala Leu Leu Leu Leu Leu Val Ala Ala Gly Arg Arg Ala 20 25 30	
50	Ala Gly Ala Ser Val Ala Thr Glu Leu Arg Cys Gln Cys Leu Gln Thr 35 40 45	
35	Leu Gln Gly Ile His Pro Lys Asn Ile Gln Ser Val Asn Val Lys Ser 50 55	
	Pro Gly Pro His Cys Ala Gln Thr Glu Val Ile Ala Thr Leu Lys Asn 65 70 75 80	
40	Gly Arg Lys Ala Cys Leu Asn Pro Ala Ser Pro Ile Val Lys Lys Ile 85 90 95	
45	Ile Glu Lys Met Leu Asn Ser Asp Lys Ser Asn 100 105	
-	(2) INFORMATION FOR SEQ ID NO:17:	
50	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 1050 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
55	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
	CTCGCCAGCT CTTCCGCTCC TCTCACAGCC GCCAGACCCG CCTGCTGAGC CCCATGGCCC	60
60	GCGCTGCTCT CTCCGCCGCC CCCAGCAATC CCCGGCTCCT GCGAGTGGCA CTGCTGCTCC	120
	TGCTCCTGGT AGCCGCTGGC CGGCGCGCAG CAGGAGCGTC CGTGGCCACT GAACTGCGCT	180
65	GCCAGTGCTT GCAGACCCTG CAGGGAATTC ACCCCAAGAA CATCCAAAGT GTGAACGTGA	240
	AGTCCCCCGG ACCCCACTGC GCCCAAACCG AAGTCATAGC CACACTCAAG AATGGGCGGA	300
	AAGCTTGCCT CAATCCTGCA TCCCCCATAG TTAAGAAAAT CATCGAAAAG ATGCTGAACA	360

	GTGACAAATC CAACTGACCA G	AGGGAGGA GGAAGCTC	AC TGGTGGCTGT	TCCTGAAGGA 420
	GGCCCTGCCC TTATAGGAAC AC	AAGAGGAA AGAGAGAC	AC AGCTGCAGAG	GCCACCTGGA 480
5	TTGTGCCTAA TGTGTTTGAG CA	TCGCTTAG GAGAAGTC	TT CTATTTATTT	ATTTATTCAT 540
	TAGTTTTGAA GATTCTATGT TA	ATATTTA GGTGTAA	AT AATTAAGGGT	ATGATTAACT 600
	CTACCTGCAC ACTGTCCTAT TA	ATATTCATT CTTTTTG#	AA TGTCAACCCC	AAGTTAGTTC 660
10	AATCTGGATT CATATTTAAT T	GAAGGTAG AATGTTTT	CA AATGTTCTCC	AGTCATTATG 720
	TTAATATTC TGAGGAGCCT GC	CAACATGCC AGCCACTG	TG ATAGAGGCTG	GCGGATCCAA 780
15	GCAAATGGCC AATGAGATCA T	GTGAAGGC AGGGGAAT	GT ATGTGCACAT	CTGTTTTGTA 840
	ACTGTTTAGA TGAATGTCAG T	GTTATTTA TTGAAAT	SAT TTCACAGTGT	GTGGTCAACA 900
20	TTTCTCATGT TGAAACTTTA AG	SAACTAAAA TGTTCTAA	AT ATCCCTTGGA	CATTTATGT 960
20	CTTTCTTGTA AGGCATACTG CO	CTTGTTTAA TGGTAGTT	TT ACAGTGTTTC	TGGCTTAGAA 1020
	CAAAGGGGCT TAATTATTGA TO	STTTTCGGA		1050
25	(2) INFORMATION FOR SEQ	ID NO:18:		
30	(B) TYPE: amin (C) STRANDEDNI (D) TOPOLOGY:	)2 amino acids no acid ESS: single linear		
25	(ii) MOLECULE TYPE:	bebride		
35	(xi) SEQUENCE DESCR	IPTION: SEO ID NO:	·18:	•
	Met Ala Arg Ala Th		÷	ro Arq Leu Leu
40	1 5		10	15
	Arg Val Ala Leu Le 20	ı Leu Leu Leu 25	Val Ala Ala S	er Arg Arg Ala 30
45	Ala Gly Ala Pro Lyd 35	3 Ala Thr Glu Lys 40	Arg Cys Gln C	ys Lys Gln Thr 5
50	Leu Gln Gly Ile Hi	Leu Lys Asn Ile 55	Gln Ser Val L	ys Val Lys Ser
50	Pro Gly Pro His Cy 65	s Ala Gln Thr Glu 70	Val Ile Ala T	hr Leu Lys Asn 80
55	Gly Gln Lys Ala Cy 85	s Leu Asn Pro Ala	Ser Pro Met V	al Lys Lys Ile 95
	Ile Glu Lys Met Le 100	l Lys		· ·
60	(2) INFORMATION FOR SEQ	ID NO:19:		
65	(i) SEQUENCE CHARA (A) LENGTH: 1 (B) TYPE: nuc (C) STRANDEDN (D) TOPOLOGY:	081 base pairs leic acid ESS: single		

-41-

	(xi) SE	QUENCE	DESCRI	PTION: S	EQ ID NO:	19:			
	CTCTCCTCCT	CGCACAC	GCCG CT	CGAACCGC	CTGCTGAG	CC CCATG	GCCCG C	GCCACGCT	60
5	TCCGCCGCCC	CCAGCAZ	ATCC CC	GGCTCCTG	CGGGTGGC	GC TGCTG	CTCCT G	CTCCTGGT	120
	GCCGCCAGCC	GGCGCGC	CAGC AG	GAGCGCCC	CTGGCCAC!	IG AACTG	CGCTG C	CAGTGCTT	180
10	CAGACCCTGC	AGGGAAT	TTCA CC	TCAAGAAC	ATCCAAAG!	rg TGAAG	STGAA G	TCCCCCGG	240
10	CCCCACTGCG	CCCAAAC	CCGA AG	TCATAGCC	ACACTCAA	GA ATGGG	CAGAA A	GCTTGTCT	300
	AACCCCGCAT	CGCCCAI	rggt ta	AGAAAATC	ATCGAAAA	GA TGCTG	AAAAA T	GGCAAATC	360
15	AACTGACCAG	AAGGAAG	GGAG GA	AGCTTATT	GGTGGCTG	IT CCTGA	AGGAG G	CCCTGCCC	r 420
	TACAGGAACA	GAAGAG	GAAA GA	GAGACACA	GCTGCAGA	GG CCACC	rggat t	GCGCCTAA	r 480
20	GTGTTTGAGC	ATCACTI	ragg ag	AAGTCTTC	TATTTATT	FA TTTAT	TATT T	ATTTGTTT	540
20	TTTTAGAAGA	TTCTATO	GTTA AT	ATTTTATG	TGTAAAAT	AA GGTTA	rgatt G	AATCTACT:	r 600
	GCACACTCTC	CCATTAI	TATT TA	TTGTTTAT	TTTAGGTC	AA ACCCAI	AGTTA G	TTCAATCC:	660
25	GATTCATATT	TAATTT	GAAG AT	AGAAGGTT	TGCAGATA	TT CTCTA	STCAT T	TGTTAATA?	720
	TTCTTCGTGA	TGACATA	ATCA CA	TGTCAGCC	ACTGTGAT	AG AGGCT	GAGGA A	rccaagaa <i>i</i>	780
30	ATGGCCAGTG	AGATCA	ATGT GA	CGGCAGGG	AAATGTAT	GT GTGTC	TTTA	GTAACTGT/	840
30	AAGATGAATG	TCAGTTO	STTA TT	TATTGAAA	TGATTTCAC	CA GTGTG	rggtc a	ACATTTCT	900
	ATGTTGAAGC	TTTAAGA	AACT AA	aatgttct	AAATATCC	CT TGGAC	ATTTT A	rgtctttc1	960
35	TGTAAGGCAT	ACTGCCI	TTGT TT	aatgttaa	TTATGCAG	rg TTTCC	CTCTG T	gttagagc <i>i</i>	1020
	GAGAGGTTTC	GATATTI	PATT GA	TGTTTTCA	CAAAGAAC	AG GAAAA	T AAAAT	ATTTAAAA	1080
40	T								1081
40	(2) INFORMA	TION FO	OR SEQ	ID NO:20	:				
45	, , , , , , , , , , , , , , , , , , ,	A) LENG B) TYPE C) STRA	STH: 10° E: amin	SS: singl	acids				
50	(ii) MO	LECULE	TYPE:	peptide					
30									
	(xi) SE	QUENCE	DESCRI	PTION: SI	EQ ID NO:2	20:			
55	Met Al	a His A	Ala Thr 5	Leu Ser	Ala Ala i	Pro Ser 1 LO	Asn Pro	Arg Leu 15	Leu
60	Arg Va	_	Leu Leu 20	Leu Leu	Leu Leu V 25	/al Ala /	Ala Ser	Arg Arg 30	Ala
50	Ala Gl	y Ala S 35	Ser Val	Val Thr	Glu Leu A 40	Arg Cys (	Sln Cys 45	Leu Gln	Thr
65	Leu Gl 50		le His	Leu Lys 55	Asn Ile (		/al Asn 50	Val Arg	Ser
	Pro Gl	y Pro H	iis Cys	Ala Gln 70	Thr Glu V	7al Ile 7 75	Ala Thr	Leu Lys	Asn 80

Gly Lys Lys Ala Cys Leu Asn Pro Ala Ser Pro Met Val Gln Lys Ile Ile Glu Lys Ile Leu Asn Lys Gly Ser Thr Asn 5 100 (2) INFORMATION FOR SEQ ID NO:21: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 988 base pairs . 10 (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 15 (x1) SEQUENCE DESCRIPTION: SEQ ID NO:21: 60 20 CCCCAGCAAT CCCCGGCTCC TGCGGGTGGC GCTGCTGCTC CTGCTCCTGG TGGCCGCCAG 120 CCGGCGCGCA GCAGGAGCGT CCGTGGTCAC TGAACTGCGC TGCCAGTGCT TGCAGACACT 180 240 GCAGGGAATT CACCTCAAGA ACATCCAAAG TGTGAATGTA AGGTCCCCCG GACCCCACTG 25 300 CGCCCAAACC GAAGTCATAG CCACACTCAA GAATGGGAAG AAAGCTTGTC TCAACCCCGC ATCCCCCATG GTTCAGAAAA TCATCGAAAA GATACTGAAC AAGGGGAGCA CCAACTGACA 360 30 GGAGAGAAGT AAGAAGCTTA TCAGCGTATC ATTGACACTT CCTGCAGGGT GGTCCCTGCC 420 CTTACCAGAG CTGAAAATGA AAAAGAGAAC AGCAGCTTTC TAGGGAOAGC TGGAAAGGAO 480 TTAATGTGTT TGACTATTTC TTACGAGGGT TCTACTTATT TATGTATTTA TTTTTGAAAG 540 CTTGTATTTT AATATTTTAC ATGCTGTTAT TTAAAGATGT GAGTGTGTTT CATCAAACAT 600 AGCTCAGTCC TGATTATTTA ATTGGAATAT GATGGGTTTT AAATGTGTCA TTAAACTAAT 660 40 ATTTAGTGGG AGACCATAAT GTGTCAGCCA CCTTGATAAA TGACAGGGTG GGGAACTGGA 720 GGGTGGGGG ATTGAAATGC AAGCAATTAG TGGATCACTG TTAGGGTAAG GGAATGTATG 780 TACACATCTA TTTTTTATAC TTTTTTTTTA AAAAAAGAAT GTCAGTTGTT ATTTATTCAA 840 45 900 ATTATCTCAC ATTATGTGTT CAACATTTTT ATGCTGAAGT TTCCCTTAGA CATTTTATGT 960 CTTGCTTGTA GGGCATAATG CCTTGTTTAA TGTCCATTCT GCAGCGTTTC TCTTTCCCTT 50 988 GGAAAAGAGA ATTTATCATT ACTGTTAC (2) INFORMATION FOR SEQ ID NO:22: (1) SEQUENCE CHARACTERISTICS: 55 (A) LENGTH: 97 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 60 (ii) MOLECULE TYPE: peptide (x1) SEQUENCE DESCRIPTION: SEQ ID NO:22: 65 Met Thr Ser Lys Leu Ala Val Ala Leu Leu Ala Ala Phe Leu Ile Ser

	Ala	Ala	Leu	Суs 20	Glu	Gly	Ala	Val	Leu 25	Pro	Arg	Ser	Ala	Lys 30	Glu	Leu	
5	Arg	Сув	Gln 35	Суз	Ile	Lys	Thr	Tyr 40	Ser	Lys	Pro	Phe	His 45	Pro	Lys	Phe	
	Ile	Lys 50	Glu	Leu	Arg	Val	Ile 55	Glu	Ser	Gly	Pro	His 60	Сув	Ala	Asn	Thr	
10	Glu 65	Ile	Ile	Val	Lys	Leu 70	Ser	Asp	Gly	Arg	Glu 75	Leu	Cys	Leu	qaA	Pro 80	
15	Lys	Glu	Asn	Trp	<b>Val</b> 85	Gln	Arg	Val	Val	Glu 90	Lys	Phe	Leu	Lys	Arg 95	Ala	
	Glu										-						
20	(2) INFO	RMAT:					_										
	(1)	(Ã (B	) LEI ) TYI ) STI	NGTH PE: 1	: 291 au <b>cl</b> e	l bas	se pa	airs									
25			) TO														
•	(xi)	SEQ	UENC	E DES	SCRII	PTIOI	N: 51	EQ II	ONO	:23:							
30	ATGACTTC																6
	GAAGGTGC	AG T	TTTG	CCAA	G GAG	STGC:	FAAA	GAAC	CTTAC	GAT (	STCA	STGC	AT A	AAGA	CATA	C	12
35	TCCAAACC	TT T	CCAC	CCCA	A ATT	TATO	CAAA	GAAG	CTGA	GAG :	rgat'	rgag	AG T	GAC	CACA	C '	18
	TGCGCCAA	CA C	AGAA	ATTA!	r TG	) AAA1	CTT	TCT	SATG	GAA (	GAGA	GCTC!	rg T	CTGG	ACCC	C	24
40	AAGGAAAA								rttt:	IGA A	AGAG	GGCT	GA G				29
	(2) INFO																
45	(1)	(B (C	UENC: ) LE: ) TY: ) ST: ) TO:	NGTH PE: 4 RAND	: 78 amino EDNE:	amin ac: SS: 4	no ao id sing:	cids									
50	(11)	MOL	ECUL	E TY	PE: 1	pept:	ide										
	(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ II	ом с	: 24 :							
55	Ala 1	Gly	Pro	Ala	Ala 5	Ala	Val	Leu	Arg	Glu 10	Lys	Arg	Cys	Val	Cys 15	Leu	
<b>6</b> 0.	Gln	Thr	Thr	Gln 20	Gly	Val	His	Pro	Lys 25	Met	Ile	Ser	Asn	Leu 30	Gln	Val.	. '
50 <sup>-</sup>	Phe	Ala	Ile 35	Gly	Pro	Gln	Сув	Ser 40	Lys	Val	Glu	Val	Val 45	Ala	Ser	Leu	
65	Lys	Asn 50	Gly	Lys	Glu	Ile	Cys 55	Leu	Asp	Pro	Glu	Ala 60	Pro	Phe	Leu	Lys	
	Lys 65	Val	Ile	Gln	Lys	Ile 70	Leu	Asp	Gly	Gly	Asn 75	Lys	Glu	Asn			

	(2) INFORMATION FOR SEQ ID NO:25:	
5	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 216 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
10		
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:	
	GTGTTGCGGG AACTGCGGTG CGTGTGTTTA CAGACCACGC AGGGAGTTCA TCCCAAAATG	60
15	ATCAGTAATC TGCAAGTGTT CGCCATAGGC CCACAGTGCT CCAAGGTGGA AGTGGTAGCC	120
	TCCCTGAAGA ACGGGAAGGA AATTTGTCTT GATCCAGAAG CCCCTTTTCT AAAGAAAGTC	180
20	ATCCAGAAAA TCCTCGACGG CGGCAACAAA GAAAAC	216
20	(2) INFORMATION FOR SEQ ID NO:26:	
25	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 93 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
30	(ii) MOLECULE TYPE: peptide	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
35	Met Gln Val Ser Thr Ala Ala Leu Ala Val Leu Leu Cys Thr Met Ala 1 5 10 15	-
40	Leu Cys Asn Gln Val Leu Ser Ala Pro Leu Ala Ala Asp Thr Pro Thr 20 25 30	
40	Ala Cys Cys Phe Ser Tyr Thr Ser Arg Gln Ile Pro Gln Asn Phe Ile 35 40 45	
45	Ala Asp Tyr Phe Glu Thr Ser Ser Gln Cys Ser Lys Pro Ser Val Ile 50 55 60	,
	Phe Leu Thr Lys Arg Gly Arg Gln Val Cys Ala Asp Pro Ser Glu Glu 65 70 75 80	
50	Trp Val Gln Lys Tyr Val Ser Asp Leu Glu Leu Ser Ala 85 90	
	(2) INFORMATION FOR SEQ ID NO:27:	
55	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 4788 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single	
60	(D) TOPOLOGY: linear	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:	
65	GAATTCAAGG CCTGTCCTGG TTTGGTCCCA ATTTACCTTT ATCATCCATA TTCACCCCCA	60
	CTGCTCTGCA GCTCCACTGA AGCACCCCCT CTTTCCTCTG AGCCACAATG TCACACCCAG	120
	GACTCTGCCT CAGCTGGGCC TCCACTGCCC ACCCATCTAT AGATGCCTAA ATCCCGGGCA	180

	GTTATCCAGA	CACAACTAAA	GTTCCATCCC	TTCCATGAAG	CCTTCCCCAA	CCCTCTGGTG	240
	GAAGGTCACT	TCTTCCTCAT	GGGGTTCTGA	GCTTTCATTT	CTTTTTCTAC	TAAGAGTTTT	300
5	ACAATTACCT	GTTCATACAC	TCTACCTGCC	CCCATGAGAC	CAGGGGCATC	TCAGAAACAA	360
	AGATCATTAA	AACCAACTAA	ATCTATTTCT	CATTATAAAA	TGAGATATGC	TGATTGATTG	420
10	CAAAATAATA	AAATAACAAA	GTATGGAAAA	GAAAAAAAAA	AGCATATAAT	CTGGCTGAGA	480
10	AGGTAGAGAC	CCTTCCACAC	CACTGAAATT	ATGTGTTGAA	AAGAATAAGG	AAAAAACTGC	540
	TTCAGTTTGG	CATTATTTAT	GTAAGTATAG	TATAGGATCC	TTAAAATGGT	TCAAAGAAAT	600
15	GGGAAATCAA	GACTTCATTT	TGGCAAAGCC	ATTGAACAGA	AACTGTAGCA	TATTTATCAG	660
	TAATTTCTTT	CAGATTAAAC	AACTGACAAC	AACCCACTTT	TCAACCAGTG	ATGTTGGAAA	720
20	TGTTTTAAAA	CAAAATTAGT	TCATAAATTT	GTGGGTTGAC	CAAGAAGGTA	ATAAAGTCTC	780
20	ACTAAATAAA	ATGAGGAAAA	TTCAGAAAAA	GAAAAAAATA	AGAAAATAAA	TCACCCATGG	840
	ATCTAAGCAC	TATTCATTCT	TTAAGGCATG	TATTTCCAAG	CCTTTTAATT	TTTTCATGCC	900
25	TAGAGTTGGC	ATGGCATATA	TATATCTTTA	TACAATTCTT	CAAATTTTAT	AGAATTTGTA	960
	TAATGTTTTA	TCTTGCTTTT	TTTTTAACCA	CTGATGTTAT	AAGCATATTT	ATGCCACTTC	1020
30	ATTCACGTTA	GAGACTTAAT	AATAAAGGAT	CTTGTGGATA	ATTTATCATT	CCCTGATAGA	1080
30	GAAAAATTTA	GCTTTGCTTA	TTTTAGAGTT	ATAAATGATG	CTGGGTCAGG	TATCTTTATG	1140
	TTTGAAGATG	GCTCCATATT	TGGGTTGTTT	CCACAGAACT	CTTTCCAGAA	ATGCTTTTTC	1200
35	TAGGTTAATG	GCTACACATA	TTTCTAGGCA	CCTGACATAC	TGACACCCAC	CTCTAAAGTA	1260
	TTTTTATGAT	CCACAACTAG	CGTTTAACAC	AGCGCCCCF/G	TCACTCCGAG	ACTAATAAAT	1320
40	AGACAAATGA	CTGAAACGTG	ACCTCATGCT	TTCTATTCCT	CCAGCTTTCA	TTGAGTTCCT	1380
₩.	TTCCTCTGGG	AGGACTGGGG	GTTGTCTAGC	CCTCCACAGC	ATCAGCCCAT	TGACCCTATC	1440
	CTTGTGGTTA	TAGCAGCTGA	GGAAGCAGAA	TTACAGCTCT	GTGGGAAGGA	ATGGGGCTGG	1500
45	AGAGTTCATG	CATAGACCAA	TTCTTTTTT	TTTTTTTTT	TGAGATGGAG	TTTCACTTTT	1560
	GTTGCCCAGG	CTGGAGTGCA	ATGGCATGAT	CTCAGCTCAC	CACAGCCCCC	ACCTCCTGGG	1620
50	TTCAAGCGAT	TCTCCTGCCC	TCAGCCTCCC	GAGTAGCTGG	GATTACAGGC	ATGTGCCACC	1680
50	ACGCCTGACT	ACTTTTGTAT	TTTTAGTAGA	GATGGAGTTT	CTCTTTCTTG	GTCAGGTTGG	1740
	TCTCAAACTC	CTGACCTCAG	GTGATCTGCA	GCCTCGGCCT	CCAAAGTGTT	GGGATTACAG	1800
55	GTGTGAGCGA	CCATGCCTGG	CTGCATAGAC	CAGTTCTTAT	GAGAAGGGAT	CAACTAAGAA	1860
	TAGCCTTGGG	TTGACACACA	CCCCTCTTCA	CACTCACAGG	AGAAACCCCA	TGAAGCTAGA	19^0
60	ACCAGTCATG	AGTTGAGAGC	TGAGAGTTAG	AGAGTAGCTC	AGAGATGCTA	TTCTTGGATA	1920
00	TCCTGAGCCC	CTGTGGTCAC	CAGGGACCCT	GAGTTGTGCA	ACACTCAGCA	TGACAGCATC	2040
	ACTACACTTA	AAAATTTCCC	TCCTCACCCC	CAGATTCCAT	TTCCCCATCC	GCCAGGGCTG	2100
65	CCTATAAAGA	GGAGAGATGG	CTTCAGACAT	CAGAAGGACG	CAGGCAGCAA	AGAGTAGTCA	2160
	GTCCCTTCTT	GGCTCTGCTG	ACACTCGAGC	CCACATTCCA	TCACCTGCTC	CCAATCATGC	2220
	AGGTCTCCAC	TGCTGCCCTT	GCCGTCCTCC	TCTGCACCAT	GGCTCTCTGC	AACCAGGTCC	2280

	TCTCTGCACC	ACGTGAGTCC	ATGTTGTTGT	TGTGGGTATC	ACCACTCTCT	GGCCATGGTT	2340
	AGACCACATC	AGTCTTTTT	TGTGGCGTGA	GAGGCCCCGA	AGAGAAAAGA	AGGAAGTTCT	2400
5	TAAAGCGCTG	CCAAACACCT	TGGTCTTTTT	CTTCACAACT	TTTATTTTTA	TCTCTAGAAG	2460
	GGGTCTTAGC	CCTCCTAGTC	TCCAGGTATG	AGAATCTAGG	CAGGGGCAGG	GGAGTTACAG	2520
	TCCCTTGTAC	AGATAGAAAA	ACAGGGTTCA	AAACGAATCA	GTTTGCAAGA	GGCAGAATCC	2580
10	AGGGCTGCTT	ACTTCCCAGT	GGGGTCTGTT	CTTCACTCTC	CAGCTCACCC	TAGTCTCCCA	2640
	GGAGCCCTGT	CCCTTGGATG	TCTTATGAGA	GATGTCCAGG	GCTTCTCTTG	GGCTGGGGTA	2700
15	TGACTTCTTG	AACCGACAAA	ATTCCATGAA	GAGAGCTAAG	AGAACAGTCC	ATTCAGGTAT	2760
	CTGGATCACA	TAGAGAAACA	GAGAACCCAC	TATGAAGAGT	CAAGGGGAAA	GAGGAATATA	2820
	GACAGAAACA	AAGAGACATT	TCTCTGCAAA	ACCCCCAAA	TGCCTTGCAG	TCACTTGGTC	2880
20	TGAGCAAGCC	TGCCCTCCTC	AACCACTCAG	GGATCAGAAG	CTGCCTGGCC	TTTTCTTCTG	2940
	AGCTGTGACT	TGGGCTTATT	CTCTCCTTTC	TCCGCAGTTG	CTGCTGACAC	GCCGACCGCC	3000
25	TGCTGCTTCA	GCTACACCTC	CCGACAGATT	CCACAGAATT	TCATAGCTGA	CTACTTTGAG	3060
	ACGAGCAGCC	AGTGCTCCAA	GCCCAGTGTC	ATGTAAGTGC	CAGTCTTCCT	GCTCACCTCT	3120
20	AGGGAGGTAG	GGAGTGTCAG	GGTGGGGGCA	GAAACAGGCC	AGAAGGCCAT	CCTGGAAAGG	3180
30	CCCAGCCTTC	AGGAGCCTAT	CGGGGATACA	GGACGCAGGG	CACTGAGGTG	TGACCTGACT	3240
	TGGGGCTGGA	GTGAGGTGGG	TGTTACAGAG	TCAGGAAGGG	CTGCCCCAGG	CCAGAGGAAA	3300
35	GGGACAGGAA	GAAGGAGGCA	GCAGGACACT	CTGAGGGCCC	CCTTGCCTGG	AGTCACTGAG	3360
	AGAAGCTCTC	TAGACGGAGA	TAGGCAGGGG	GCCCCTGAGA	GAGGAGCAGG	CCTTGAGCTG	3420
40	CCCAGGACAG	AGAGCAGGAT	GTCAGGGCCA	TGGTGGGCCC	AGGATTCCCC	GGCTGGATTC	3480
40	CCCAGTGCTT	AACTCTTCCT	CCCTTCTCCA	CAGCTTCCTA	ACCAAGAGAG	GCCGGCAGGT	3540
	CTGTGCTGAC	CCCAGTGAGG	AGTGGGTCCA	GAAATACGTC	AGTGACCTGG	AGCTGAGTGC	3600
45	CTGAGGGGTC	CAGAAGCTTC	GAGGCCCAGC	GACCTCAGTG	GGCCCAGTGG	GGAGGAGCAG	3660
	GAGCCTGAGC	CTTGGGAACA	TGCGTGTGAC	CTCCACAGCT	ACCTCTTCTA	TGGACTGGTT	3720
50	ATTGCCAAAC	AGCCACACTG	TGGGACTCTT	CTTAACTTAA	ATTTTAATTT	ATTTATACTA	3780
50	TTTAGTTTTT	ATAATTTATT	TTTGATTTCA	CAGTGTGTTT	GTGATTGTTT	GCTCTGAGAG	3840
	TTCCCCCTGT	CCCCTCCACC	TTCCCTCACA	GTGTGTCTGG	TGACAACCGA	GTGGCTGTCA	3900
55	TCGGCCTGTG	TAGGCAGTCA	TGGCACCAAA	GCCACCAGAC	TGACAAATGT	GTATCAGATG	3960
	CTTTTGTTCA	GGGCTGTGAT	CGGCCTGGGG	AAATAATAAA	GATGTTCTTT	TAAACGGTAA	4020
60	ACCAGTATTG	AGTTTGGTTT	TGTTTTTCTG	GCAAATCAAA	ATCACTGGTT	AAGAGGAATC	4080
00	ATAGGCAAAG	ATTAGGAAGA	GGTGAAATGG	AGGGAAATTG	GGAGAGATGG	GGAGCGCTGC	4140
	GACAGAGTTA	TCCACTTCAC	AAAATTCTGG	AACATTGAAA	CTACGAATAT	GTTATAACTC	4200
65	AAATCGTAAT	ATGCACGCTC	TAGGAGAATT	AACTACTTGA	ATGGCCACCA	TTAAGCAGAG	4260
	TATTCTGTAG	GGCATATTCA	TGATGAATCA	AGCTCTTAAT	AGCAATTATT	TACATTGTTG	4320
	AGGCTTACTC	CTCCTACTGA	GTGCTTTTTA	TACATTGTTC	ATTTAATCTT	ACCAATGCAA	4380

PCT/US94/08207

	TAGTACAGCT TAGGTACTAT TAATACCTCC ACTTGACAGA AAAGTAACCC AGGGCTCAGA	4440
	AAGGTTAGAC AACTTGGCTG AGGTTACACA GCACGTAAAC GGTCAATTGT GTTCCAAAAC	4500
5	TGGACTTTTA TTGAACTACA GACTATGCTG TTAACCATTG ACCAAGTTAT TTCCCAAAGT	4560
	ATGACCCGCC TATACTCAAA TCTTACCCCA TTCTTTAACA GATGATACTT TATCCATTGC	4620
10	AACCACTTCC TGTCAGGATT CTGAGTTGAC ATAGAGTGTT TCAGCAGTGA TTATTTAAGC	4680
10	CAATTACATC AGGATCTTTA GGTGTAGACC TGGGAACTGA TATTTTTATC AAGCTCATGA	4740
	GGTGTTCCAT AGCATGTTAA TGACTGAGAG CCACTGTCAA TAGAATTC	4788
15	(2) INFORMATION FOR SEQ ID NO:28:	
20	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 92 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
25		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:	
30	Met Lys Leu Cys Val Thr Val Leu Ser Leu Leu Met Leu Val Ala Ala 1 5 10 15	
	Phe Cys Ser Pro Ala Leu Ser Ala Pro Met Gly Ser Asp Pro Pro Thr 20 25 30	
35	Ala Cys Cys Phe Ser Tyr Thr Ala Arg Lys Leu Pro Arg Asn Phe Val 35 40 45	
	Val Asp Tyr Tyr Glu Thr Ser Ser Leu Cys Ser Gln Pro Ala Val Val 50 60	
40	Phe Gln Thr Lys Arg Ser Lys Gln Val Cys Ala Asp Pro Ser Glu Ser	
	65 70 75 80	
45	Trp Val Gln Glu Tyr Val Tyr Asp Leu Glu Leu Asn 85	
	(2) INFORMATION FOR SEQ ID NO:29:	
50	(1) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 696 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
55		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:	
60	TTCCCCCCCC CCCCCCCCC CCCCCCCGA GCACAGGACA CAGCTGGGTT CTGAAGCTTC	60
υυ	TGAGTTCTGC AGCCTCACCT CTGAGAAAAC CTCTTTTCCA CCAATACCAT GAAGCTCTGC	120
	GTGACTGTCC TGTCTCCCT CATGCTAGTA GCTGCCTTCT GCTCTCCAGC GCTCTCAGCA	180
65	CCAATGGGCT CAGACCCTCC CACCGCCTGC TGCTTTTCTT ACACCGCGAG GAAGCTTCCT	240
	CGCAACTTTG TGGTAGATTA CTATGAGACC AGCAGCCTCT GCTCCCAGCC AGCTGTGGTA	300
	TTCCAAACCA AAAGAAGCAA GCAAGTCTGT GCTGATCCCA GTGAATCCTG GGTCCAGGAG	360

WO 95/04158 PCT/US94/08207

-49-

	GGGCTCTGGA AACCACATGG CTTCACCTGT CCCCGAAACT ACCAGCCCTA CACCATTCCT	420
	TCTGCCCTGC TTTTGCTAGG TCACAGAGGA TCTGCTTGGT CTTGATAAGC TATGTTGTTG	480
5	CACTTTAAAC ATTTAAATTA TACAATCATC AACCCCCAAC	520
	(2) INFORMATION FOR SEQ ID NO:32:	
10	(1) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 99 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
15	(ii) MOLECULE TYPE: peptide	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:	
	Met Lys Val Ser Ala Ala Leu Leu Cys Leu Leu Leu Ile Ala Ala Thr 1 10 15	
25	Phe Ile Pro Gln Gly Lys Ala Gln Pro Asp Ala Ile Asn Ala Pro Val 20 25 30	
	Thr Cys Cys Tyr Asn Phe Thr Asn Arg Lys Ile Ser Val Gln Arg Leu 35 40	
30	Ala Ser Tyr Arg Arg Ile Thr Ser Ser Lys Cys Pro Lys Glu Ala Val 50 60	
35	Ile Phe Lys Thr Ile Val Ala Lys Glu Ile Cys Ala Asp Pro Lys Gln 65 70 75 80	
	Lys Trp Val Gln Asp Ser Met Asp His Leu Asp Lys Gln Thr Gln Thr 85 90 95	
40	Pro Lys Thr	
	(2) INFORMATION FOR SEQ ID NO:33:	
45	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 725 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
50		
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:33:	
55	CTAACCCAGA AACATCCAAT TCTCAAACTG AAGCTCGCAC TCTCGCCTCC AGCATGAAAG	60
JJ	TCTCTGCCGC CCTTCTGTGC CTGCTGCTCA TAGCAGCCAC CTTCATTCCC CAAGGGCTCG	120
	GTGAGCEAGA TGCAATCAAT GCGCCAGTCA CGTGGTGTTA TAACTTCAGC AATAGGAAGA	180
60	TCTCAGTGCA GAGGCTCGCG AGCTATAGAA GAATCACCAG CAGCAAGTGT CCCAAAGAAG	240
	CTGTGATCTT CAAGACCATT GTGGCCAAGG AGATCTGTGC TGACCCCAAG CAGAAGTGGG	300
65	TTCAGGATTC CATGGACCAC CTGGACAAGC AAACCCAAAC TCCGAAGACT TGAACACTCA	360
	CTCCACAACC CAAGAATCTG CAGCTAACTT ATTTTCCCCT AGCTTTCCCC AGACACCCTG	420
	TTTTATTTTA TTATAATGAA TTTTGTTTGT TGATGTGAAA CATTATGCCT TAAGTAATGT	480

WO 95/04158 PCT/US94/08207

-50-

	TAATTCTTAT TTAAGTTATT GATGTTTTAA GTTTATCTTT CATGGTACTA GTGTTTTTA	34
	GATACAGAGA CTTGGGGAAA TTGCTTTTCC TCTTGAACCA CAGTTCTACC CCTGGGATGT	60
5	TTTGAGGGTC TTTGCAAGAA TCATTAATAC AAAGAATTTT TTTTAACATT CCAATGCATT	66
	GCTAAAATAT TATTGTGGAA ATGAATATTT TGTAACTATT ACACCAAATA AATATATTTT	72
10	TGTAC	72
10	(2) INFORMATION FOR SEQ ID NO:34:	
15	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 99 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
20 ·	(11) MOLECULE TYPE: peptide	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:34:	
25	Met Lys Ala Ser Ala Ala Leu Leu Cys Leu Leu Leu Thr Ala Ala Ala 1 5 10 15	
30	Phe Ser Pro Gln Gly Leu Ala Gln Pro Val Gly Ile Asn Thr Ser Thr 20 25 30	
,,,	Thr Cys Cys Tyr Arg Phe Ile Asn Lys Lys Ile Pro Lys Gln Arg Leu 35 40 45	
35	Glu Ser Tyr Arg Arg Thr Thr Ser Ser His Cys Pro Arg Glu Ala Val 50 55 60	
	Ile Phe Lys Thr Lys Leu Asp Lys Glu Ile Cys Ala Asp Pro Thr Gln 65 70 75 80	
10	Lys Trp Val Gln Asp Phe Met Lys His Leu Asp Lys Lys Thr Gln Thr 85 90 95	
	Pro Lys Leu	
15	(2) INFORMATION FOR SEQ ID NO:35;	
50	(1) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 810 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	٠
55	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:35:	
	AGCAGAGGGG CTGAGACCAA ACCAGAAACC TCCAATTCTC ATGTGGAAGC CCATGCCCTC	60
60	ACCCTCCAAC ATGAAAGCCT CTGCAGCACT TCTGTGTCTG CTGCTCACAG CAGCTGCTTT	120
	CAGCCCCCAG GGGCTTGCTC AGCCAGTTGG GATTAATACT TCAACTACCT GCTGCTACAG	180
	ATTTATCAAT AAGAAAATCC CTAAGCAGAG GCTGGAGAGC TACAGAAGGA CCACCAGTAG	240
55	CCACTGTCCC CGGGAAGCTG TAATCTTCAA GACCAAACTG GACAAGGAGA TCTGTGCTGA	300
	CCCCACACAG AAGTGGGTCC AGGACTTTAT GAAGCACCTG GACAAGAAAA CCCAAACTCC	360

	AAAGCTTTGA ACATTCATGA CTGAACTAAA AACAAGCCAT GACTTGAGAA ACAAATAATT	420
	TGTATACCCT GTCCTTTCTC AGAGTGGTTC TGAGATTATT TTAATCTAAT TCTAA GGAAT	480
5	ATGAGCTTTA TGTAATAATG TGAATCATGG TTTTTCTTAG TAGATTTTAA AAGTTATTAA	540
	TATTTTAATT TAATCTTCCA TGGATTTTGG TGGGTTTTGA ACATAAAGCC TTGGATGTAT	600
10	ATGTCATCTC AGTGCTGTAA AAACTGTGGG ATGCTCCTCC CTTCTCTACC TCATGGGGGT	660
10	ATTGTATAAG TCCTTGCAAG AATCAGTGCA AAGATTTGCT TTAATTGTTA AGATATGATG	720
	TCCCTATGGA AGCATATTGT TATTATATA TTACATATTT GCATATGTAT GACTCCCAAA	780
15	TTTTCACATA AAATAGATTT TTGTAAAAAA	810
	(2) INFORMATION FOR SEQ ID NO:36:	
20	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 91 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
25	(ii) MOLECULE TYPE: peptide	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:	
	Met Lys Val Ser Ala Ala Arg Leu Ala Val Ile Leu Ile Ala Thr Ala 1 10 15	
35	Leu Cys Ala Pro Ala Ser Ala Ser Pro Tyr Ser Ser Asp Thr Thr Pro 20 25 30	•
	Cys Cys Phe Ala Tyr Ile Ala Arg Pro Leu Pro Arg Ala His Ile Lys 35 40 45	
40	Glu Tyr Phe Tyr Thr Ser Gly Lys Cys Ser Asn Pro Ala Val Val Phe 50 60	
45	Val Thr Arg Lys Asn Arg Gln Val Cys Ala Asn Pro Glu Lys Lys Trp 65 70 75 80	
45	Val Arg Glu Tyr Ile Asn Ser Leu Glu Met Ser 85 90	
<b>.</b>	(2) INFORMATION FOR SEQ ID NO:37:	
50	(1) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 1160 base pairs (B) TYPE: nucleic acid	
55	(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:	
	CCTCCGACAG CCTCTCCACA GGTACCATGA AGGTCTCCGC GGCACGCCTC GCTGTCATCC	60
	TCATTGCTAC TGCCCTCTGC GCTCCTGCAT CTGCCTCCCC ATATTCCTCG GACACCACAC	120
55	CCTGCTGCTT TGCCTACATT GCCCGCCCAC TGCCCCGTGC CCACATCAAG GAGTATTTCT	180
	ACACCAGTGG CAAGTGCTCC AACCCAGCAG TCGTCTTTGT CACCCGAAAG AACCGCCAAG	240
	TGTGTGCCAA CCCAGAGAAG AAATGGGTTC GGGAGTACAT CAACTCTTTG GAGATGAGCT	300

	AGGATGGAGA GTCCTTGAAC CTGAACTTAC ACAAATTTGC CTGTTTCTGC TTGCTCTTGT	36
	CCTAGCTTGG GAGGCTTCCC CTCACTATCC TACCCCACCC GCTCCTTGAA GGGCCCAGAT	42
5	TCTGACCACG ACGAGCAGCA GTTACAAAAA CCTTCCCCAG GCTGGACGTG GTGGCTCAGC	48
	CTTGTAATCC CAGCACTTTG GGAGGCCAAG GTGGGTGGAT CACTTGAGGT CAGGAGTTCG	54
10	AGACAGCCTG GCCAACATGA TGAAACCCCA TGTGTACTAA AAATACAAAA AATTAGCCGG	60
, 10	GCGTGGTAGC GGGCGCCTGT AGTCCCAGCT ACTCGGGAGG CTGAGGCAGG AGAATGGCGT	66
	GAACCCGGGA GCGGAGCTTG CAGTGAGCCG AGATCGCGCC ACTGCACTCC AGCCTGGGCG	72
15	ACAGAGCGAG ACTCCGTCTC AAAAAAAAAA AAAAAAAAA AAAAAATACA AAAATTAGCC	78
	GCGTGGTGGC CCACGCCTGT AATCCCAGCT ACTCGGGAGG CTAAGGCAGG AAAATTGTTT	84
20	GAACCCAGGA GGTGGAGGCT GCAGTGAGCT GAGATTGTGC CACCTCACTC CAGCCTGGGT	90
20	GACAAAGTGA GACTCCGTCA CAACAACAAC AACAAAAAGC TTCCCCAACT AAAGCCTAGA	96
	AGAGCTTCTG AGGCGCTGCT TTGTCAAAAG GAAGTCTCTA GGTTCTGAGC TCTGGCTTTG	102
25	CCTTGGCTTT GCAAGGGCTC TGTGACAAGG AAGGAAGTCA GCATGCCTCT AGAGGCAAGG	1080
	AAGGGAGGAA CACTGCACTC TTAAGCTTCC GCCGTCTCAA CCCCTCACAG GAGCTTACTG	1140
30	GCAAACATGA AAAATCGGGG	1160
	(2) INFORMATION FOR SEQ ID NO:38:	
35	(1) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 97 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
40	(ii) MOLECULE TYPE: peptide	
	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:38:	
45	Met Arg Ile Ser Ala Thr Leu Leu Cys Leu Leu Leu Ile Ala Ala 10 15	
50	Phe Ser Ile Gln Val Trp Ala Gln Pro Asp Gly Pro Asn Ala Ser Thr 20 25 30	
	Cys Cys Tyr Val Lys Lys Gln Lys Ile Pro Lys Arg Asn Leu Lys Ser 35 40 45	
55	Tyr Arg Arg Ile Thr Ser Ser Arg Cys Pro Trp Glu Ala Val Ile Phe 50 60	
	Lys Thr Lys Lys Gly Met Glu Val Cys Arg Glu Ala His Gln Lys Trp 65 75 80	
60	Val Glu Glu Ala Ile Ala Tyr Leu Asp Met Lys Thr Pro Thr Pro Lys	

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(2) INFORMATION FOR SEQ ID NO:39:

5	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 593 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
. 10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:	
	ACTGAAGCCA GCTCTCTCAC TCTCTTTCTC CACCATGAGG ATCTCTGCCA CGCTTCTGTG	60
15	CCTGCTGCTC ATAGCCGCTG CTTTCAGCAT CCAAGTGTGG GCCCAACCAG ATGGGCCCAA	120
	TGCATCCACA TGCTGCTATG TCAAGAACE AAGATCCCC AAGAGGAATC TCAAGAGCTA	180
. DO	CAGAAGGATC ACCAGTAGTC GGTGTCCCTG GGAAGCTGTT ATCTTCAAGA CAAAGAAGGG	240
··20	CATGGAAGTC TGTCGTGAAG CCCATCAGAA GTGGGTCGAG GAGGCTATAG CATACTTAGA	300
	CATGAAAACC CCAACTCCAA AGCCTTGAAG AAATGTGCCT GAACAGAAAC CAACCTAGGA	360
25	GCCAAGAAGC AAAAATTCCT CACCGCTGTT CTTTCTGAGA ACTGTTGATG AAATGTGTTG	420
	ATCACGGTCC TAAGGGATAG GAGCTGTCTG TAGGAATGTG AAACAGTCAC GCCTAAGGAA	480
20	TGGTCTTTAA GTTATTAATA TTTTTATTTA ATTAGCCATG TACTTTGGTG TGATTTGAAT	540
30	GTAAAGCTCT GGAGACCTCA TGTCACTTTA ACATTGTGTT AGCTGCAGAA TTC	593
	(2) INFORMATION FOR SEQ ID NO:40:	
35	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 72 amino acids	
	(B) TYPE: amino acid (C) STRANDEDNESS: single	
40	(D) TOPOLOGY: linear	
	(11) MOLECULE TYPE: peptide	
45		
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:	
	Asp Ser Val Ser Ile Phe Ile Thr Cys Cys Phe Asn Val Ile Asn Arg 1 5 10 15	
50	Lys Ile Pro Ile Gln Arg Leu Glu Ser Tyr Thr Arg Ile Thr Asn Ile 20 25 30	
55	Gln Cys Pro Lys Glu Ala Val Ile Phe Lys Thr Gly Lys Glu Val Cys 35 40 45	
33	Ala Asp Pro Lys Glu Arg Trp Val Arg Asp Ser Met Lys His Lys Asp 50 55 60	
60	Gln Ile Phe Gln Asn Leu Lys Pro 65 70	
	(2) INFORMATION FOR SEQ ID NO:41:	
65	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 15 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	

	(11) MOLECULE TYPE: peptide	
5	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:	
	Asn Leu Ala Lys Gly Lys Glu Glu Ser Leu Asp Ser Asp Leu Cys 1 5 10 15	
10	(2) INFORMATION FOR SEQ ID NO:42:	
15	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 32 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: peptide	
20		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:	
25	Cys Asn Gln Val Glu Val Ile Ala Thr Leu Lys Asp Gly Arg Lys Ile 1 5 10 15	
	Cys Leu Asp Pro Asp Ala Pro Arg Ile Lys Lys Ile Val Gln Lys Lys 20 25 30	
30	(2) INFORMATION FOR SEQ ID NO:43:	
35	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 96 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
10	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:43:	
-	T001100110 T00110 T0011	<i>-</i> ^
		60
15	SILICINGIN INITITION MAMMA	96

#### **CLAIMS**

#### We claim:

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- 1. A method of screening for AHA compounds comprising the steps of:
  - a) contacting a compound with radiolabeled heparin/heparan sulfate and heparanase;
  - maintaining the compounds in contact with the radiolabeled heparin/heparan sulfate and heparanase for a time and under such conditions sufficient to allow inhibition of heparanase activity;
  - c) detecting inhibition of heparanase activity (a compound that gives 50% inhibition at a concentration of 1 µM or less); and
  - d) selecting compounds that inhibit heparanase activity.
- 2. A method according to claim 1 wherein the heparanase is recombinant.
- 15 3. A heparanase having an isoelectric point of less than 5.5 and possessing activity greater than 20 units heparanase activity per µg protein.
  - 4. A heparanase according to Claim 3, having an isoelectric point of about 4.8 5.1.
- 5. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions and activated with transglutaminase, having an amino acid sequence selected from the group consisting of SEQ. ID NO: 1, SEQ. ID NO: 3, SEQ. ID NO: 5 or SEQ. ID NO: 7.
- 25 6. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions, having an amino acid sequence of SEQ, ID NO; 1.
  - 7. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions and activated with transglutaminase, having an amino acid sequence of SEQ. ID NO: 3.
  - 8. A heparanase according to Claim 4, purified to apparent homogeneity, prepared in the presence of reducing conditions and activated with transglutaminase.
- 35 9. A heparanase, as in claim 3, prepared by recombinant means, activated with transglutaminase and having an amino acid sequence selected from the group consisting of SEQ.

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ID NO: 1, SEQ. ID NO: 3, SEQ. ID NO: 5 or SEQ. ID NO: 7.

- 10. A heparanase, as in claim 3, prepared by recombinant means, activated with transglutaminase and having an amino acid sequence of SEQ. ID NO: 1.
- 11. A heparanase, as in claim 3, prepared by recombinant means, activated with transglutaminase and having an amino acid sequence of SEQ. ID NO: 3.
- 12. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID. NO: 12, SEQ. ID. NO: 14; SEQ. ID. NO: 16, SEQ. ID. NO: 18, SEQ. ID. NO: 20, SEQ. ID. NO: 22 and SEQ. ID. NO: 24.
- 13. A heparanase purified to apparent homogeneity, as in claim 3, prepared in the presence
  15 of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID. NO: 26, SEQ. ID. NO: 28, SEQ. ID. NO: 30,
  SEQ. ID. NO: 32; SEQ. ID. NO: 34, SEQ. ID. NO: 36, SEQ. ID. NO: 38 and SEQ.ID. NO: 40.
- 14. A method according to claim 1 wherein the heparanase is purified to apparent homogeneity, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID. NO: 12, SEQ. ID. NO: 14; SEQ. ID. NO: 16, SEQ. ID. NO: 18, SEQ. ID. NO: 20, SEQ. ID. NO: 22 and SEQ. ID. NO: 24.
- 25 15. A method according to claim 1 wherein the heparanase is purified to apparent homogeneity, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID. NO: 26, SEQ. ID. NO: 28, SEQ. ID. NO: 30, SEQ. ID. NO: 32; SEQ. ID. NO: 34, SEQ. ID. NO: 36, SEQ. ID. NO: 38 and SEQ. ID. NO: 40.
  - 16. A method according to claim 1 wherein the heparanase is purified to apparent homogeneity, prepared in the presence of reducing conditions and activated with transglutaminase, and having an amino acid sequence selected from the group consisting of SEQ. ID NO: 1, SEQ. ID NO: 3, SEQ. ID NO: 5 or SEQ. ID NO: 7.
  - 17. A peptide having an amino acid sequence of SEQ. ID. NO: 42.

## INTERNATIONAL SEARCH REPORT

Inter nal Application No
PCT/US 94/08207

A CLAS	SIEICATION OF SUID-		PCT/US 94/08207
ÎPC 6	SSIFICATION OF SUBJECT MATTER C12Q1/34 C12N9/24 C12N9	/96 C07K14/4	7
According	to International Patent Classification (IPC) or to both national	classification and IPC	
B. FIELD	DS SEARCHED		
IPC 6	documentation searched (classification system followed by class C12Q C12N A61K C07K	ification symbols)	
Document	ation searched other than minimum documentation to the extent	that such documents are include	ed in the fields searched
Electronic	data base consulted during the international scarcb (name of data	a base and, where practical, see	rch terms use."
C. DOCUM	MENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the	he relevant passages	Relevant to claim No.
A	ANALYTICAL BIOCHEMISTRY, vol.157, no.1, 15 August 1986,	NEW YORK US	1,2
	pages 162 - 171 MOTOWO NAKAJIMA ET AL. 'A Solid Substrate of Heparanase: Its Ap to Assay of Human Melanoma for Sulfate Degradative Activity' see the whole document	I-Phase	
A	JOURNAL OF BIOLOGICAL CHEMISTRY vol.259, no.4, 25 February 1984 BALTIMORE, MD US pages 2283 - 2290 MOTOWO NAKAJIMA ET AL. 'Metasta	•	1,2
<b>A</b>	Melanoma Cell Heparanase' see the whole document see page 2289, left column, lin- right column, line 31	e 63 -	3,4
Y Purth	er documents are listed in the continuation of box C.	-/	
		X Patent family mem	bers are listed in annex.
A" document consider	egories of cited documents : nt defining the general state of the art which is not red to be of particular relevance	cited to understand the	d after the international filing date t in conflict with the application but principle by theory underlying the
L' documen which is	nt which may throw doubts on priority claim(s) or a cited to establish the multication date of another	involve an inventive st	relevance; the claimed invention ovel or cannot be considered to up when the document is taken alone
O' document other ma	or other special reason (as specified)  no referring to an oral disclosure, use, exhibition or  eans	document is combined ments, such combination	relevance; the claimed invention involve an inventive step when the with one or more other such docu- in being obvious to a person skilled
	at published prior to the international filing date but in the priority date claimed	in the art. "&" document member of the	
	ctual completion of the international search		sternational search report
	December 1994	2 1. 12. 9	
lame and ma	ulling address of the ISA  European Patent Office, P.B. 5818 Patentiaan 2  NL - 2280 HV Rijswijk  Td. (+ 31-70) 340-2040, Tx. 31 651 epo nl,  Fax (+31-70) 340-3016	Authorized officer  Döpfer, K-	P

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Inta onal Application No PCT/US 94/08207

C/Continue	Dion) DOCUMENTS CONSIDERED TO BE RELEVANT	PCT/US 94/08207		
Category *	Citation of document, with indication, where appropriate, of the relevant passages			
	and managed, where appropriate, or the relevant passages	Relevant to claim No.		
<b>A</b>	WO,A,91 02977 (HADASSAH MEDICAL ORGANIZATION) 7 March 1991 cited in the application see page 23, paragraph 6.1.5 - page 26, paragraph 6.1.7; claims 1-3	3-16		
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information on patent family members

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